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U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

217039US0XPCT

U.S. APPLICATION NO. (IF KNOWN - SEE 37 CFR

09/926799

INTERNATIONAL APPLICATION NO.
PCT/JP00/04095INTERNATIONAL FILING DATE
22 JUNE 2000PRIORITY DATE CLAIMED
22 JUNE 1999

TITLE OF INVENTION

SRSV DETECTION KIT

APPLICANT(S) FOR DO/EO/US

Naokazu TAKEDA, et al.

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (24) indicated below.
4. ☒ The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
 - a. ☐ is attached hereto (required only if not communicated by the International Bureau).
 - b. ☒ has been communicated by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
 - a. ☒ is attached hereto.
 - b. ☐ has been previously submitted under 35 U.S.C. 154(d)(4).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
 - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ have been communicated by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
10. ☐ An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).
11. ☒ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☒ A copy of the International Search Report (PCT/ISA/210).

Items 13 to 20 below concern document(s) or information included:

13. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☐ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☐ A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.
20. ☐ A second copy of the published international application under 35 U.S.C. 154(d)(4).
21. ☐ A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
22. ☐ Certificate of Mailing by Express Mail
23. ☒ Other items or information:

Notice of Priority / PCT/IB/304 / PCT/IB/308
PTO-1449 / Drawings (6 sheets)

U.S. APPLICATION NO. (IF KNOWN) SEE 37 CFR 1.53 09/928799		INTERNATIONAL APPLICATION NO. PCT/JP00/04095		ATTORNEY'S DOCKET NUMBER 217039US0XPCT	
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24. The following fees are submitted:

BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :	CALCULATIONS	PTO USE ONLY																																																																	
<input type="checkbox"/> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1040.00																																																																			
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Surcharge of \$130.00 for furnishing the oath or declaration later than _____ months from the earliest claimed priority date (37 CFR 1.492 (e)). <input type="checkbox"/> 20 <input type="checkbox"/> 30	\$0.00																																																																		
<table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="width: 15%;">CLAIMS</th> <th style="width: 20%;">NUMBER FILED</th> <th style="width: 20%;">NUMBER EXTRA</th> <th style="width: 10%;">RATE</th> <th style="width: 35%;"></th> </tr> </thead> <tbody> <tr> <td>Total claims</td> <td>14 - 20 =</td> <td>0</td> <td>x \$18.00</td> <td style="text-align: right;">\$0.00</td> </tr> <tr> <td>Independent claims</td> <td>7 - 3 =</td> <td>4</td> <td>x \$84.00</td> <td style="text-align: right;">\$336.00</td> </tr> <tr> <td colspan="4">Multiple Dependent Claims (check if applicable). <input checked="" type="checkbox"/></td> <td style="text-align: right;">\$280.00</td> </tr> <tr> <td colspan="4" style="text-align: right;">TOTAL OF ABOVE CALCULATIONS =</td> <td style="text-align: right;">\$1,506.00</td> </tr> <tr> <td colspan="4"> <input type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2. </td> <td style="text-align: right;">\$0.00</td> </tr> <tr> <td colspan="4" style="text-align: right;">SUBTOTAL =</td> <td style="text-align: right;">\$1,506.00</td> </tr> <tr> <td colspan="4"> Processing fee of \$130.00 for furnishing the English translation later than _____ months from the earliest claimed priority date (37 CFR 1.492 (f)). <input type="checkbox"/> 20 <input type="checkbox"/> 30 </td> <td style="text-align: right;">\$0.00</td> </tr> <tr> <td colspan="4" style="text-align: right;">TOTAL NATIONAL FEE =</td> <td style="text-align: right;">\$1,506.00</td> </tr> <tr> <td colspan="4"> Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable). <input type="checkbox"/> </td> <td style="text-align: right;">\$0.00</td> </tr> <tr> <td colspan="4" style="text-align: right;">TOTAL FEES ENCLOSED =</td> <td style="text-align: right;">\$1,506.00</td> </tr> <tr> <td colspan="4"></td> <td style="text-align: right;">Amount to be: refunded \$</td> </tr> <tr> <td colspan="4"></td> <td style="text-align: right;">charged \$</td> </tr> </tbody> </table>	CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		Total claims	14 - 20 =	0	x \$18.00	\$0.00	Independent claims	7 - 3 =	4	x \$84.00	\$336.00	Multiple Dependent Claims (check if applicable). <input checked="" type="checkbox"/>				\$280.00	TOTAL OF ABOVE CALCULATIONS =				\$1,506.00	<input type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.				\$0.00	SUBTOTAL =				\$1,506.00	Processing fee of \$130.00 for furnishing the English translation later than _____ months from the earliest claimed priority date (37 CFR 1.492 (f)). <input type="checkbox"/> 20 <input type="checkbox"/> 30				\$0.00	TOTAL NATIONAL FEE =				\$1,506.00	Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable). <input type="checkbox"/>				\$0.00	TOTAL FEES ENCLOSED =				\$1,506.00					Amount to be: refunded \$					charged \$		
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a. ☒ A check in the amount of **\$1,506.00** to cover the above fees is enclosed.


b. ☐ Please charge my Deposit Account No. _____ in the amount of _____ to cover the above fees. A duplicate copy of this sheet is enclosed.

c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. **15-0030**. A duplicate copy of this sheet is enclosed.

d. ☐ Fees are to be charged to a credit card. **WARNING:** Information on this form may become public. Credit card information should not be included on this form. Provide credit card information and authorization on PTO-2038.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:



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REGISTRATION NUMBER

Dec. 20 2001

DATE

DESCRIPTION

SRSV DETECTION KIT

5 Technical Field

This invention relates to a kit for detecting and distinguishing one or more small round structure viruses (hereinafter called "SRSVs") in a specimen.

Background Art

10 SRSVs are a group of causative viruses of human viral gastroenteritis, the discovery of the first one of which goes back to 1972. They are known to cause infantile acute gastroenteritis and also outbreaks of food poisoning or the like among adults and preschool or elementary school children. Due
15 to the inability to proliferate these SRSVs by cell culture and the lack of animal models capable of exhibiting sensitivity thereto, SRSV antigens and anti-SRSV antibodies are hardly available, resulting in a delay in the development of immunoserologic methods for the detection of the viruses.

20 Under such circumstances, it was succeeded to clone the gene of the Norwalk virus, an SRSV, in 1993, leading to the determination of the base sequence of its complete genomes [JP(PCT) 6-506823 A]. Subsequently, PCR methods which are useful to amplify a part of an RNA polymerase region were developed,
25 and 14 SRSV-related viruses have been found to date. As a result

of analyses of about 120 amino acids in these RNA polymerase regions, SRSVs are considered to be roughly differentiated into two genogroups, that is, Genogroup I including the Norwalk virus strain as a prototype and Genogroup II including the Snow Mountain virus strain as a prototype.

As genetic analyses of SRSV-related viruses proceeded, it came to knowledge that substantial diversity exists even in the same genogroup. As a matter of fact, it was found that with an RT-PCR method making use of primers for the genes of the Norwalk virus and Snow Mountain virus strains as the prototypes of the respective genogroups, every SRSV is not detectable and also that it is very difficult to design primers or set RT-PCT conditions for achieving efficient amplification of SRSVs.

In the meantime, antigens were prepared against some of the viruses, such as the Norwalk virus strain and the Snow Mountain strain, by genetic expression, antibodies were obtained, and ELISA-dependent SRSV detection methods making use of such antibodies were also developed. It was, however, still impossible to detect every gastroenteritis-causing SRSV due to the diversity of the SRSVs.

In Japan, on the other hand, SRSVs were designated in 1997 to be causative factors of food poisoning as defined in the Food Sanitation Act so that, if SRSV food poisoning breaks out, determination of its infection route is required. There is accordingly a desire for a method which easily and surely detects

and identifies SRSVs in infected subjects' feces or foods.

Disclosure of the Invention

Accordingly, an object of the present invention is to
5 provide a kit which can easily detect from a specimen an
SRSV-related virus known to date and can surely discriminate
its serotype and genogroup.

With the foregoing circumstances in view, the present
inventors have proceeded with an genetic and immunological
10 investigation on SRSV-related viruses. As a result, it has been
found that combined use of antibodies obtained from 11
SRSV-related virus peptides, including newly-found novel virus
peptides, can detect most SRSVs in specimens and can surely
discriminate the serotypes and genogroups of the SRSVs, leading
15 to the completion of the present invention.

Specifically, the present invention provides an SRSV
detection kit comprising all antibodies against SRSV-related
virus constituting peptides selected from the following peptide
groups (a) to (k), respectively:

20 (a) a peptide having an amino acid sequence represented
by SEQ ID NO: 1 and peptides each having at least 80% of homology
with said amino acid sequence, and partial peptides thereof,

(b) a peptide having an amino acid sequence represented
by SEQ ID NO: 2 and peptides each having at least 80% of homology
25 with said amino acid sequence, and partial peptides thereof,

(c) a peptide having an amino acid sequence represented by SEQ ID NO: 3 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

5 (d) a peptide having an amino acid sequence represented by SEQ ID NO: 4 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(e) a peptide having an amino acid sequence represented by SEQ ID NO: 5 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

10 (f) a peptide having an amino acid sequence represented by SEQ ID NO: 6 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(g) a peptide having an amino acid sequence represented by SEQ ID NO: 7 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(h) a peptide having an amino acid sequence represented by SEQ ID NO: 8 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

20 (i) a peptide having an amino acid sequence represented by SEQ ID NO: 9 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(j) a peptide having an amino acid sequence represented by SEQ ID NO: 10 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

25 and

(k) a peptide having an amino acid sequence represented by SEQ ID NO: 11 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof.

The present invention also provides an SRSV detection kit for discriminating SRSVs in genogroup, said SRSV detection kit comprising all antibodies against SRSV-related virus constituting peptides selected from the following peptide groups (a) to (d), respectively:

(a) a peptide having an amino acid sequence represented by SEQ ID NO: 1 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(b) a peptide having an amino acid sequence represented by SEQ ID NO: 2 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(c) a peptide having an amino acid sequence represented by SEQ ID NO: 3 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof, and

(d) a peptide having an amino acid sequence represented by SEQ ID NO: 4 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof.

Further, the present invention also provides an SRSV detection kit for discriminating genogroup of SRSVs, said SRSV detection kit comprising all antibodies against SRSV-related virus constituting peptides selected from the following peptide

groups (e) to (k), respectively:

(e) a peptide having an amino acid sequence represented by SEQ ID NO: 5 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

5 (f) a peptide having an amino acid sequence represented by SEQ ID NO: 6 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(g) a peptide having an amino acid sequence represented by SEQ ID NO: 7 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

10 (h) a peptide having an amino acid sequence represented by SEQ ID NO: 8 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(i) a peptide having an amino acid sequence represented by SEQ ID NO: 9 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(j) a peptide having an amino acid sequence represented by SEQ ID NO: 10 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

20 and

(k) a peptide having an amino acid sequence represented by SEQ ID NO: 11 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof.

Furthermore, the present invention also provides
25 SRSV-related virus strain genes having base sequences

represented by SEQ ID NOS: 15, 20, 21 and 22 or base sequences similar to the first-mentioned base sequences, respectively, except for deletion, replacement or addition of one to several bases of said first-mentioned base sequences.

5

Brief Description of the Drawings

FIG. 1 is an electron micrograph (x 100,00) of virus-like particles derived from the Hu/NLV/Seto 124/1989/JP strain.

10 FIG. 2 is an electron micrograph (x 100,00) of virus-like particles derived from the Hu/NLV/Funabashi 258/1996/JP strain.

FIG. 3 is an electron micrograph (x 100,00) of virus-like particles derived from the Hu/NLV/Chiba 407/1987/JP strain.

FIG. 4 is an electron micrograph (x 100,00) of virus-like particles derived from the Hu/NLV/Narita 104/1997/JP strain.

15 FIG. 5 is an electron micrograph (x 100,00) of virus-like particles derived from the Hu/NLV/Sanbu 809/1998/JP strain.

FIG. 6 is an electron micrograph (x 100,00) of virus-like particles derived from the Hu/NLV/Ichikawa 754/1998/JP strain.

20 FIG. 7 is an electron micrograph (x 100,00) of virus-like particles derived from the Hu/NLV/Chitta 1876/1996/JP strain.

FIG. 8 is an electron micrograph (x 100,00) of virus-like particles derived from the Hu/NLV/Kashiwa 47/1997/JP strain.

FIG. 9 is an electron micrograph (x 100,00) of virus-like particles derived from the Hu/NLV/Mie 7k/1994/JP strain.

25 FIG. 10 is an electron micrograph (x 100,00) of virus-like

particles derived from the Hu/NLV/Kashiwa 645/1999/JP strain.

FIG. 11 is an electron micrograph (x 100,00) of virus-like particles derived from the Hu/NLV/Osaka 10-25/1999/JP strain.

5 **Best Modes for Carrying Out the Invention**

1. SRSV-related viruses

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The SRSV detection kit according to the present invention is characterized by the use of the antibodies against SRSV-related virus constituting peptides having the 11 specific amino acid sequences or at least 80% of homologies with the amino acid sequences in the groups (a) to (k). Of these, the peptides belonging to the group (d), the group (i), the group (j) and the group (k) are novel peptides different from any SRSV-related viruses registered with the GeneBank to date (Table 1, which will be described subsequently herein). Owing to the incorporation of the 11 antibodies, including antibodies against these novel peptides, into the kit, SRSV-related viruses can be detected without omission.

20 The SRSV-related virus constituting peptides useful in the present invention embrace their mutants in each of which one or more amino acids have been deleted from, replaced in or added to its corresponding amino acid sequence; and also their mutants in each of which one or several bases have been deleted from, replaced in or added to a base sequence encoding its
25 corresponding amino acid sequence.

Illustrative of the peptide having the amino acid sequence represented by the SEQ ID NO: 1 in the group (a) is a virus constituting peptide of the Hu/NLV/Kashiwa 645/1999/JP strain obtained from feces of an SRSV infected patient in Japan, whereas
5 examples of the peptides each having at least 80% of homology with the amino acid sequence include one derived from the Desert Shield/90/SA strain (GeneBank Accession No. U04469).

Illustrative of the peptide having the amino acid sequence represented by the SEQ ID NO: 2 in the group (b) is a virus
10 constituting peptide of the Hu/NLV/Seto 124/1989/JP strain obtained from feces of an SRSV infected patient in Japan, whereas examples of the peptides each having at least 80% of homology with the amino acid sequence include those derived from the KY-89/89J strain (GeneBank Accession No. L23828) and the
15 Norwalk/68/US strain (GeneBank Accession No. M876611).

Illustrative of the peptide having the amino acid sequence represented by the SEQ ID NO: 3 in the group (c) is a virus constituting peptide of the Hu/NLV/Funabashi 258/1996/JP strain obtained from feces of an SRSV infected patient in Japan, whereas
20 examples of the peptides each having at least 80% of homology with the amino acid sequence include one derived from the Southampton/91/UK strain (GeneBank Accession No. L07418).

Illustrative of the peptide having the amino acid sequence represented by the SEQ ID NO: 4 in the group (d) is a virus
25 constituting peptide of the Hu/NLV/Chiba 407/1987/JP strain

obtained from feces of an SRSV infected patient in Japan.

The peptide having the amino acid sequence represented by SEQ ID NO: 4 has less than 75% of homology in structural gene (SEQ ID NO: 15) with any one of the SRSV-related virus strains (Table 1, which will be described subsequently herein) registered with the GeneBank to date, and is a peptide having a novel sequence not reported to date.

Illustrative of the peptide having the amino acid sequence represented by the SEQ ID NO: 5 in the group (e) is a virus constituting peptide of the Hu/NLV/Narita 104/1997/JP strain obtained from feces of an SRSV infected patient in Japan, whereas examples of the peptides each having at least 80% of homology with the amino acid sequence include those derived from the Bristol/93/UK strain (GeneBank Accession No. X76716), the Lordsdale/93/UK strain (GeneBank Accession No. X86557), and the Camberwell/94/AU strain (GeneBank Accession No. U46500).

Illustrative of the peptide having the amino acid sequence represented by the SEQ ID NO: 6 in the group (f) is a virus constituting peptide of the Hu/NLV/Sanbu 809/1998/JP strain obtained from feces of an SRSV infected patient in Japan, whereas examples of the peptides each having at least 80% of homology with the amino acid sequence include those derived from the Mexico/89/MEX strain (GeneBank Accession No. U22498), the Auckland strain (GeneBank Accession No. U460391), the Toronto/77/CA strain (GeneBank Accession No. U02030), and the

OTH-25/89/J strain (GeneBank Accession No. L23830).

Illustrative of the peptide having the amino acid sequence represented by the SEQ ID NO: 7 in the group (g) is a virus constituting peptide of the Hu/NLV/Ichikawa 754/1998/JP strain obtained from feces of an SRSV infected patient in Japan, whereas examples of the peptides each having at least 80% of homology with the amino acid sequence include those derived from the Snow Mountain/76/US strain (GeneBank Accession No. U70059) and the Melksham/89/UK strain (GeneBank Accession No. X81879).

Illustrative of the peptide having the amino acid sequence represented by the SEQ ID NO: 8 in the group (h) is a virus constituting peptide of the Hu/NLV/Chitta 1876/1996/JP strain obtained from feces of an SRSV infected patient in Japan, whereas examples of the peptides each having at least 80% of homology with the amino acid sequence include one derived from the Hawaii/71/US strain (GeneBank Accession No. U07611).

Illustrative of the peptide having the amino acid sequence represented by the SEQ ID NO: 9 in the group (i) is a virus constituting peptide of the Hu/NLV/Kashiwa 47/1997/JP strain obtained from feces of an SRSV infected patient in Japan.

The peptide having the amino acid sequence represented by SEQ ID NO: 9 has less than 75% of homology in structural gene (SEQ ID NO: 20) with any one of the SRSV-related virus strains (Table 1, which will be described subsequently herein) registered with the GeneBank to date, and is a peptide having a novel sequence

not reported to date.

Illustrative of the peptide having the amino acid sequence represented by the SEQ ID NO: 10 in the group (j) is a virus constituting peptide of the Hu/NLV/Mie 7k/1994/JP strain
5 obtained from feces of an SRSV infected patient in Japan.

The peptide having the amino acid sequence represented by SEQ ID NO: 10 has less than 70% of homology in structural gene (SEQ ID NO: 21) with any one of the SRSV-related virus strains (Table 1, which will be described subsequently herein) registered
10 with the GeneBank to date, and is a peptide having a novel sequence not reported to date.

Illustrative of the peptide having the amino acid sequence represented by the SEQ ID NO: 11 in the group (k) is a virus constituting peptide of the Hu/NLV/Osaka 10-25/1999/JP strain
15 obtained from feces of an SRSV infected patient in Japan.

The peptide having the amino acid sequence represented by SEQ ID NO: 11 has less than 70% of homology in structural gene (SEQ ID NO: 22) with any one of the SRSV-related virus strains (Table 1, which will be described subsequently herein) registered
20 with the GeneBank to date, and is a peptide having a novel sequence not reported to date.

Table 1

Virus strain	GeneBank Accession No.
Desert Shield/90/SA	U04469
Norwalk/68/US	M876611
KY-89/89J	L23828
OTH-25/89/J	L23830
Southampton/91/UK	L07418
Lordsdale/93/UK	X86557
Bristol/93/UK	X76716
Camberwell/94/AU	U46500
Toronto/77/CA	U02030
Mexico/89/MEX	U22498
Snow Mountain/76/US	U70059
Melksham/89/UK	X81879
Auckland	U460391
Hawaii/71/US	U07611

The SRSV-related virus constituting peptides in these groups (a) to (k) embrace, in addition to the above-described peptides, partial peptides each of which contains a specific amino acid sequence in its corresponding peptide and has antigenecity equivalent to the corresponding peptide.

According to a homological analysis of about 120 amino acids of RNA polymerase regions of the SRSV-related virus constituting peptides, these SRSV-related virus constituting peptides can be classified into two genogroups. Described

specifically, they can be classified into Type I to which the peptides in the groups (a) to (d) belong and Type II to which the peptides in the groups (e) to (k) belong.

2. Cloning of the SRSV-related virus constituting genes

From feces of an SRSV infected patient, viral RNA is extracted using the cetyltrimethylammonium bromide (CTAB) method or the like, cDNA was formed by an oligo-dT primer and a reverse transcriptase, and using the cDNA and primers capable of amplifying structural gene regions of the individual SRSV-associate viruses, PCR was conducted to amplify structural gene fragments.

Such a structural gene fragment is inserted in a plasmid by once conducting TA cloning with an *E. coli* cloning vector.

As a cloning vector usable here, it is possible to use a known cloning vector such as a vector derived from a plasmid obtained using as host procaryotic cells represented by *E. coli* or from a bacteriophage represented by ϕ phage, and appropriately combined use of a cloning vector and its host cell is desired. Specific examples of the cloning vector include pBR322, pUC19 and pCRII. The insertion of the DNA can be conducted by a method known *per se* in the art, and upon formation of such a vector, use of *E. coli* cells is desired as they permit easy genetic manipulation.

3. Expression of structural gene and creation of virus-like particles.

By having fragments of the above-obtained individual virus constituting genes in the groups (a) to (k) expressed with a suitable expression system or by using virus-like particles created from the virus constituting peptides in a genetic engineering manner, antibodies against the respective viruses can be obtained. A description will hereinafter be made about an expression when *E. coli* is used and also about the creation of virus-like particles.

(1) Expression by *E. coli*

Plasmids with the structural gene regions of the respective SRSV-related viruses incorporated therein, respectively, are each digested with a restriction endonuclease which does not cleave the structural gene region. Then, the structural gene region is collected and incorporated, for example, in pGEX (GST fusion protein expression vector; product of Pharmacia AB), pTrc99A (*E. coli* expression vector; product of Pharmacia AB), pTrxFus (thioredoxin fusion protein expression vector; product of Invitrogen Corporation), pET (expression vector making use of pT7RNA promoter; product of Novagen Inc.), a maltose binding protein expression vector, or a α galactosidase fusion protein expression vector. At this time, the structural gene region to be incorporated can be of the complete length or can be a partial region, with a partial region containing at least one antigen epitope of an SRSV being preferred. Gene expression vectors with the structural gene regions incorporated therein

as described above are transformed by an *E coli* strain suited for gene expression, for example, the BL21 strain, the DH10B strain, the JM109 strain or the XL1-Blue strain. Expression of the gene can be conducted by culturing the thus-obtained transformants in a general liquid culture medium, for example, L-broth. It is preferred for the expression to add a gene expression promoter, for example, IPTG or, when a PL promoter is used, to apply a heat shock.

Purification of a peptide so expressed can be conducted following a general purification method for expressed protein, which makes use of *E coli*. If the expressed protein is in a dissolved form, for example, its purification can be conducted by affinity chromatography making use of a GST column or a column for maltose binding proteins. If the expressed protein is in an insoluble form, its purification can be achieved by conducting affinity chromatography making use of a Ni chelate.

(2) Creation of SRSV virus-like particles

A plasmid with a structural gene region of an SRSV-related virus incorporated therein is digested with a restriction endonuclease which does not cleave the structural gene region. Then, the structural gene region is collected and incorporated, for example, in a baculovirus transfer vector such as pVL1393. The transfer vector and a linear baculovirus DNA, from which a gene region essential for proliferation has been deleted, are subjected to transfection in insect cells such that homologous

recombination is induced to form the target recombinant baculovirus.

By infecting the thus-obtained recombinant baculovirus to insect cells such as Sf9 cells or Tn5 cells and incubating the infected insect cells under adequate growth conditions in a manner known *per se* in the art, the structural protein of the SRSV is expressed. By allowing the structural protein to undergo self-assembly, virus-like particles can be produced. Use of a biochemical purification method, for example, centrifugation makes it possible to isolate and purify the virus-like particles. Whether or not such virus-like particles have been formed can be confirmed by subjecting the self-assembled product to negative staining with uranyl acetate and examining the stained self-assembled product by an electron microscope.

The virus-like particles obtained as described above do not have infectiveness as they do not contain any gene internally. Nonetheless, they have antigenicity equivalent to virus particles because they structurally have substantially the same shape as virus particles.

4. Acquisition of antibodies against SRSV-related viruses

By immunizing an animal with the thus-obtained virus constituting peptide or virus-like particles, an anti-SRSV-related virus antibody can be prepared. Incidentally, such an antibody can be either a monoclonal antibody or a polyclonal antibody.

Preparation of an immune antibody by making use of virus-like particles can be conducted, for example, as will be described next. In a manner known *per se* in the art, a rabbit is immunized with virus-like particles of one of the SRSV-related viruses, and from separated serum, an IgG antibody (anti-SRSV antibody) against the virus-like particles can be obtained. For the separation and isolation of the antibody, a method such as DEAE Sepharose chromatography can be used.

Using the 11 types of virus-like particles of the groups (a) to (k) obtained as described above and their corresponding anti-SRSV antibodies, their cross reactivities were measured. As will be shown below in Table 2, absolutely no cross-reactivity was exhibited between the individual SRSV-related viruses. According to the SRSV detection method of the present invention, it is therefore possible to concurrently discriminate the serotypes of 11 types of SRSVs. This also indicates the possibility of discriminating Genogroup I Genogroup II from each other at the same time.

5. Detection of SRSV-related viruses

For the detection of one or more SRSVs in a specimen by the individual anti-SRSV antibodies obtained as described, conventionally employed immunoassays making use of antigen-antibody reactions, for example, radioimmunoassay by the sandwich technique, enzyme-linked immunosorbent assay (ELISA) and the like can be used, with ELISA being particularly

preferred. Described specifically, the 11 types of anti-SRSV antibodies are separately poured into a microplate to prepare an SRSV screening plate. A dilution of a fecal emulsion, which has been prepared from feces of an SRSV infected patient, is added to the wells of the plate, and is then allowed to react. Peroxidase (POD) labeled anti-SRSV antibodies of the respective viruses are thereafter added and reacted. After a substrate solution (TMB containing hydrogen peroxide) is added and reacted, 0.6 N sulfuric acid is added to stop the reactions. By measuring the absorbance (450 nm/630 nm) of each well by an ELISA autoreader, the SRSV or SRSVs can be detected.

When it is desired to conduct only the detection of one or more SRSVs in a specimen, a detection kit can be prepared by using a microplate with all the 11 types of anti-SRSV antibodies mixed and immobilized thereon. To also discriminate even the serotypes of the one or more SRSVs, a detection kit can be prepared by using microplates with all the 11 types of anti-SRSV antibodies immobilized separately thereon.

Further, the discrimination of the genogroups is feasible by a kit making use of a microplate with antibodies against the peptides in the groups (a) to (d) mixed and immobilized thereon (Type I plate) or a microplate with antibodies against the peptides in the groups (e) to (k) mixed and immobilized thereon (Type II plate).

Moreover, immobilization of the individual anti-SRSV

antibodies useful in the present invention with a carrier such as a latex or magnetic beads makes it possible to surely capture one or more SRSV-related viruses in a specimen. The carrier with one or more SRSV-associate viruses captured thereon can be recovered by centrifugation in the case of the latex or by magnet in the case of the magnetic beads. Subsequent to the recovery, virus RNAs can be extracted and used.

Examples

The SRSV detection kits according to the present invention will hereinafter be specifically described based on Examples.

Example 1 Cloning of structural genes of SRSV-related viruses

(1) Synthesis of cDNA

PBS (9 mL) and "Daiflon" (1 mL) were added to feces (0.5 to 1.0 g) of an SRSV patient, followed by homogenization. The homogenate was then centrifuged at 3,000 rpm for 20 minutes, and the supernatant was collected as a 10% fecal emulsion.

Using a 1-mL aliquot of the fecal emulsion, RNA of the SRSV was extracted by the cetyltrimethylammonium bromide (CTAB) method, and the RNA was eventually suspended in a 0.1% diethyl pyrocarbonate solution (30 μ L). Using the suspension, cDNA was prepared by a reverse transcriptase derived from the Oligo-dT(12-18) primer and AMV (Avian Myeloblastosis Virus) (product of SEIKAGAKU CORPORATION).

(2) Isolation of structural gene regions

Using the cDNA prepared in (1) and primers for amplifying the structural gene regions shown below, PCR was conducted. Subsequent to the PCR, amplified structural gene fragments were separated by agarose gel electrophoresis, and were then recovered by using "SuprecTM-01" (TAKARA).

Hu/NLV/Kashiwa 645/1999/JP gene:G1/F2(SEQ ID NO: 23),
Oligo-dT(33) (SEQ ID NO: 24)

Hu/NLV/Seto 124/1989/JP gene:G1/F2(SEQ ID NO: 23),G1/R0
(SEQ ID NO: 25)

Hu/NLV/Funabashi 258/1996/JP gene:G1/F2(SEQ ID NO: 23),
Oligo-dT(33)(SEQ ID NO: 24)

Hu/NLV/Chiba 407/1987/JP gene:D5(SEQ ID NO: 26),
CV-U4(SEQ ID NO: 27)

Hu/NLV/Narita 104/1997/JP gene:97k104/F1(SEQ ID NO: 28),
97k104/R1(SEQ ID NO: 29)

Hu/NLV/Sanbu 809/1998/JP gene:G2/F3(SEQ ID NO: 30),
MV-R1(SEQ ID NO: 31)

Hu/NLV/Ichikawa 754/1998/JP gene:G2/F3(SEQ ID NO: 30),
SMV-R1(SEQ ID NO: 32)

Hu/NLV/Chitta 1876/1996/JP gene:G2/F3(SEQ ID NO: 30),
G2/R0(SEQ ID NO: 33)

Hu/NLV/Kashiwa 47/1997/JP gene:97k104/F1(SEQ ID NO: 28),
Oligo-dT(33)(SEQ ID NO: 24)

Hu/NLV/Mie 7k/1994/JP gene:G2/F3(SEQ ID NO: 30),
Oligo-dT(33)(SEQ ID NO: 24)

Hu/NLV/Osaka 10-25/1999/JP gene:GFCR7(SEQ ID NO: 34),
Oligo-dT(33)(SEQ ID NO: 24)

(3) Cloning of structural genes

TA cloning of the recovered structural gene fragments to
an *E. coli* cloning vector, pCRII(product of Invitrogen
Corporation) was conducted. Obtained from these clones were
plasmids with the structural genes of the viruses incorporated
therein, pCRII/645, pCRII/124, pCRII/258, pCRII/Chiba,
pCRII/104, pCRII/809, pCRII/754, pCRII/76, pCRII/47, pCRII/7k,
and pCRII/10-25.

Example 2 Determination of base sequences

Determination of the base sequences of the structural genes
of the Hu/NLV/Kashiwa 645/1999/JP strain, the Hu/NLV/Seto
124/1989/JP strain, the Hu/NLV/Funabashi 258/1996/JP strain,
the Hu/NLV/Chiba 407/1987/JP strain, the Hu/NLV/Narita
104/1997/JP strain, Hu/NLV/Sanbu 809/1998/JP strain, the
Hu/NLV/Ichikawa 754/1998/JP strain, the Hu/NLV/Chitta
1876/1996/JP strain, the Hu/NLV/Kashiwa 47/1997/JP strain, the
Hu/NLV/Mie 7k/1994/JP strain, and the Hu/NLV/Osaka
10-25/1999/JP strain was conducted in the below-described
manner.

Firstly, a primer (first primer) was set in the vicinity
of the polyhedrin promoter of pVL1393 as a transfer vector, and
by the dye termination method, a labeling reaction was conducted
by using a "Cycle Sequencing Kit FS" (product of Perkin-Elmer

Corp.). The DNA concentration of the transfer vector employed was 0.4 $\mu\text{g}/\mu\text{L}$, whereas the concentration of the sequencing primer used was 3.2 $\text{pmol}/\mu\text{L}$. Subsequent to the reaction, the excess fluorescent pigment was eliminated using a centrprep spin column (manufactured by Perkin-Elmer Corp.). The reaction mixture was completely dried by a vacuum lyophilizer, and the lyophilizate was suspended in a special sample buffer (20 μL ; product of Perkin-Elmer Corp.). Subsequent to stirring, the suspension was subjected to centrifugal precipitation. The precipitate was dried at 95°C for 2 minutes. After quenching, it was analyzed by an autosequencer ("ABI Genetic Analyzer 310").

Using the base sequence determined by the first primer, a new sequencing primer (second primer) was set on the 3'-side of the base sequence. Using this second primer, a labeling reaction was conducted by a cyclic sequencing kit in a similar manner as mentioned above. Subsequent to the reaction, operation similar to that mentioned above was performed, and the base sequence was analyzed by the autosequencer. As has been described above, a sequencing primer was set on the 3' side of the base sequence determined in each cycle, and determination of the base sequence was conducted. By repeating this procedure, the base sequences from the 5'-ends to the 3'-ends of the 11 types of SRSV-related virus structural genes (SEQ ID NO: 12 to SEQ ID NO 22) were determined. Among these, the base sequences represented by SEQ ID NO: 15 (the Hu/NLV/Chiba 407/1987/JP

strain), SEQ ID NO: 20 (the Hu/NLV/Kashiwa 47/1997/JP strain),
SEQ ID NO: 21 (the Hu/NLV/Mie 7k/1994/JP strain) and SEQ ID
NO: 22 (the Hu/NLV/Osaka 10-25/1999/JP strain) were confirmed
to be novel sequences not reported to date.

5 Example 3 Creation of recombinant baculovirus capable of
 yielding virus-like particles

 (1) Construction of transfer vectors

 The plasmids with the structural gene regions incorporated
therein, which had been obtained in Example 1(3), were digested
10 by a restriction endonuclease which does not cleave the
structural gene regions. Subsequent to separation by agarose
gel electrophoresis, the structural gene regions were recovered
by "SuprecTM01" (TAKARA). The recovered gene fragments were
incorporated in baculovirus transfer vectors pVL1393 (product
15 of Invitrogen Corporation), which had been digested by the same
restriction endonuclease, to prepare transfer vectors.

 (2) Creation of recombinant baculoviruses

 Baculovirus DNA (0.5 μ g; "Baculo-Gold") and one of the
transfer vectors (1 μ g) obtained in (1) were dissolved in
20 distilled water (8 μ L). The resulting solution was mixed with
a two-fold dilution of lipofectin (equivalent amount), and the
thus-obtained mixture was left over at room temperature for 15
minutes. After Sf9 cells (1×10^5 cells) suspended in an insect
cell culture medium, "Ex-cell 400", were adsorbed at 26.5°C for
25 30 minutes in a plastic Petri dish (diameter: 3.5 cm), a mixture

of the transfer vector and "Baculo-Gold" was added dropwise to the cells, followed by incubation at 26.5°C. 24 Hours later, the culture medium was replaced by a "TC100" (product of GIBCO BRL Life Technologies; hereinafter referred to as "TC100") which
5 contained 10% fetal bovine serum and 2% BTB (products of GIBCO BRL Life Technologies), and incubation was continued further.

(3) Purification of recombinant baculoviruses

After each recombinant baculovirus obtained in (2) was incubated for 5 days, the culture supernatant was diluted tenfold
10 with an insect cell culture medium such as TC100. A 0.1-mL aliquot of the diluted supernatant was taken, and inoculated to 3×10^6 Sf9 cells cultured in a plastic Petri dish of 3.5 cm in diameter. Subsequent to adsorption at 26.5°C for 60 minutes, TC100 culture medium (2 mL) which contained 1% of Agarose ME
15 (low melting-point agarose) was overlayed, followed by incubation at 26.5°C. On the 4th day after the initiation of the incubation, TC100 (1 mL) which contained 0.005% of neutral red was further overlayed, followed by incubation at 26.5°C. On the following day, the formed plaques were scraped off with
20 a microtip. and were suspended in TC100 culture medium.

(4) Production of recombinant baculovirus seeds and measurement of their infective potencies

Each suspension obtained in (3) was inoculated to 1×10^7 Sf9 cells. Subsequent to adsorption at 26.5°C for 60 minutes,
25 TC100 was added, followed by incubation at 26.5°C for 3 to 4

days. The culture was centrifuged at 2,500 rpm for 10 minutes at 4°C, and the culture supernatant was collected. The collected culture supernatant was inoculated to 1×10^7 Sf9 cells. Subsequent to adsorption at 26.5°C for 60 minutes, TC100 was added, followed by incubation at 26.5°C for 3 to 4 days.

Next, the culture supernatant was inoculated to 3×10^7 Sf9 cells cultured in a plastic Petri dish of 3.5 cm in diameter. Subsequent to adsorption at 26.5°C for 60 minutes, TC100 culture medium (2 mL) which contained 1% of Agarose ME (low melting-point agarose) was overlayed, followed by incubation at 26.5°C. On the 4th day after the initiation of the incubation, TC100 (1 mL) which contained 0.005% of neutral red was then overlayed, followed by incubation at 26.5°C. On the following day, the formed plaques were measured to calculate the infective potency of the recombinant baculovirus. This was recorded as the infective potency of the recombinant baculovirus.

Example 4 Creation of virus-like particles

(1) Expression of structural proteins by using recombinant baculoviruses

Each recombinant baculovirus was infected at M.O.I.s (Multiplicities of infection) of 1 to 10 to Sf9 insect cells. Upon infection, a suspension of the recombinant baculovirus was added dropwise to the cells, and the recombinant baculovirus was subjected to adsorption for about 60 minutes or so with gentle shaking. After that, TC100 was added as an insect cell culture

medium, followed by incubation at 26.5°C for 5 to 6 days.

(2) Identification of expressed proteins

The culture supernatant of each recombinant virus infection was periodically sampled. After having been resolved
5 by SDS-PAGE, the protein was detected by Coomassie blue staining, and by an expected molecular weight, the validity of the expressed protein was confirmation. Further, subsequent to resolving the protein by SDS-PAGE, the protein was transferred onto a nitrocellulose membrane, and by the Western blotting technique,
10 the expressed protein was then identified with a convalescent serum of the SRSV.

(3) Purification and recovery of virus-like particles

The recombinant baculovirus seeds were infected at M.O.I.s of from 1 to 10. Subsequent to adsorption for about 60 minutes,
15 "Ex-cell 400" was added, followed by incubation at 26.5°C for 3 days. A protease inhibitor, for example, pepstatin A or a leupeptin, was then added to the culture to a final concentration of 1 mM, followed by further incubation for 2 to 3 days.

Subsequent to the incubation, the culture was
20 centrifuged at 2,500 rpm for 10 minutes at 4°C to collect the culture supernatant. The collected culture was centrifuged at 10,000 rpm for 30 minutes to eliminate the recombinant baculovirus. The supernatant was centrifuged at 25,000 rpm for 4 hours on a "Beckmann SW28 Rotor" to have virus-like particles
25 precipitated. Then, the centrifuge tube from which the

supernatant had been discarded was held upside down to complete eliminate the supernatant. After that, Grace buffer or PBS(-) (0.5 mL) with the protease inhibitor added therein was added to the centrifuge tube, and the centrifuge was allowed to stand overnight at 4°C.

After the standing, the virus-like particles were suspended in the protease-inhibitor-containing Grace buffer which had been added, and were recovered. To the recovered virus-like particles, protease-inhibitor-containing Grace buffer or PBS(-) with CsCl (3.8 g) added therein was added to give 13 mL. The resulting mixture was ultracentrifuged at 16°C and 35,000 rpm for 24 to 48 hours. Subsequent to the ultracentrifugation, a pale band in which virus-like particles gathered was collected. After 5-fold dilution with protease-inhibitor-containing Grace buffer, the resultant suspension was ultracentrifuged at 45,000 rpm for 3 hours on a "Beckmann TL100.3 Rotor" to have the virus-like particles precipitated.

The precipitated virus-like particles were solubilized with Grace buffer or PBS(-) to which the protease inhibitor had been added. Protease-inhibitor-containing Grace buffer solutions which contained 10% to 50% of sucrose were prepared in a 4PA tube, into which the solubilized solution of the virus-like particles was overlayed, followed by sucrose density-gradient centrifugation at 35,000 rpm for 4 hours at

4°C. Subsequent to the centrifugation, a pale band of virus-like particles was collected as purified SRSV virus-like particles in a 1-mL syringe fitted with a 26G needle.

The purified SRSV virus-like particles was diluted with Grace buffer as needed, and the quantity of protein was measured by the Bradford method.

The purified SRSV virus-like particles were subjected to negative staining with uranyl acetate, and were then examined by an electron microscope to ascertain whether or not virus-like particles had been formed (FIGS. 2 to 12).

Example 5 Preparation of Immune Antibodies and Labeled by Use of Antibodies Virus-like Particles

(1) Preparation of immune antibodies against virus-like particles

A phosphate buffer (pH 7.2, 1 mL) - which contained the purified SRSV virus-like particles (500 ig) obtained from one of the Hu/NLV/Kashiwa 645/1999/JP strain, the Hu/NLV/Seto 124/1989/JP strain, the Hu/NLV/Funabashi 258/1996/JP strain, the Hu/NLV/Chiba 407/1987/JP strain, the Hu/NLV/Narita 104/1997/JP strain, Hu/NLV/Sanbu 809/1998/JP strain, the Hu/NLV/Ichikawa 754/1998/JP strain, the Hu/NLV/Chitta 1876/1996/JP strain, the Hu/NLV/Kashiwa 47/1997/JP strain, the Hu/NLV/Mie 7k/1994/JP strain, and the Hu/NLV/Osaka 10-25/1999/JP strain - and the Freund's incomplete adjuvant (1 mL) were mixed, and then immunized to a New Zealand white rabbit

(3 kg) in a manner known *per se* in the art. Three weeks later, the rabbit was immunized further with a mixture of a phosphate buffer (pH 7.2, 1 mL), which contained the SRSV virus-like particles (0.25 ig), and the Freund's incomplete adjuvant (1 mL) (booster dose). Additional 3 weeks later, immunization was conducted as in the booster dose, and about 7 to 10 days after the additional booster dose, exsanguination was conducted, and the serum component was separated.

After the separated and purified serum was subjected to ammonium sulfate fractionation, the relevant fraction was dialyzed overnight at 4°C against 50 mM Tris-HCl (pH 7.6). The inner dialyzate was then subjected to DEAE Sepharose chromatography which had been equilibrated with 50 mM Tris-HCl (pH 7.6). Under monitoring at an UV wavelength of 280 nm, an O.D. peak was collected to obtain an DEAE-purified IgG antibody (anti-SRSV antibody) against the virus-like particles.

(2) Preparation of labeled antibodies

Each anti-SRSV antibody was labeled with POD by an improved periodic acid technique ["Koso Men-eki Sokuteiho (Enzyme Immunoassay)", 2, 91, 1982]. Described specifically, POD was dissolved at 4 mg/mL in distilled water and 0.1 M sodium periodate (0.2 mL) was added, followed by a reaction at room temperature for about 20 minutes. The reaction mixture was then dialyzed overnight against 1 mM sodium acetate buffer (pH 4.0).

Subsequent to the dialysis, 0.2 M sodium carbonate buffer (pH

9.5, 0.02 mL) was added to adjust the pH to 9.5, and at the same time, the anti-SRSV antibody (8 mg) was added.

After having been allowed to react at room temperature for 2 hours, 4 mg/mL sodium borohydroxide (0.1 mL) was added, followed by a reaction at 4°C for about 2 hours. After the reaction, gel filtration was conducted with "Sephacryl S-200" while using 10 mM phosphate buffer. Under monitoring at an UV wavelength of 280 nm, a POD-labeled anti-SRSV antibody fraction was collected.

(3) Preparation of a solid-phase anti-SRSV antibody microplate

The anti-SRSV antibodies were separately diluted with a carbonate buffer (pH 9.5) to a concentration of from 0.5 to 10 µg/mL and then poured at 100 µL/well into a polystyrene flat-bottom microplate (manufactured by Nunc). The microplate was then allowed to stand overnight 4°C. After standing for 18 hours or longer, the microplate was washed 3 to 4 times at 200 µL/well with PBS which contained "Tween 20" at a final concentration of 0.05%. 10 mM PBS (pH 7.2) - which contained bovine serum albumin (BSA) and "Tween 20" at final concentrations of 0.5% and 0.05%, respectively - was then added at 200 µL/well. The microplate was allowed to stand overnight 4°C to obtain a solid-phase anti-SRSV antibody microplate.

Example 6 Cross-reactivity

(1) Antigen detection ELISA

The purified SRSV virus-like particles of each group were diluted to 4 ng/mL to 0.04 ng/mL with a solution containing bovine serum albumin (BSA) and "Tween 20" at final concentrations of 0.2% and 0.05%, respectively, in a buffer (10 mM PBS, pH 7.2).

5 Then, the diluted emulsions of the virus-like particles (VLPs) were each added at 100 μ L/well to wells of the corresponding solid-phase anti-SRSV antibody microplate, followed by a reaction at room temperature for 60 minutes. After the reaction, the reaction mixtures in the wells were eliminated under suction.

10 10 mM PBS (pH 7.2) which contained "Tween 20" at a final concentration of 0.05% was added at 200 μ L/well to the wells, and was then eliminated under suction likewise. This procedure was repeated at least three times. After washing, the POD-labeled anti-SRSV antibody of the corresponding serotype,

15 which had been diluted 20000-fold with a buffer, was added at 100 μ L/well, followed by a reaction at room temperature for 60 minutes. Subsequent to washing, a TMB solution with hydrogen peroxide contained therein was added at 100 μ L/well, followed by a reaction at room temperature for 30 minutes. After the

20 reaction, 0.6 N sulfuric acid was added at 100 μ L/well, and the absorbance (450 nm/630 nm) of each well was measured by an ELISA autoreader. The results are shown in Table 2.

Table 2 Cross-reactivity between Serotypes

Purified VLP	VLP concentration (ng/mL)	Solid-phase antibody plate x POD (top: strain name, bottom: dilution of POD-labeled antibody)													7k	10-25
		124	258	407	645	104	809	754	1876	47	20000	20000	20000	20000		
124	4	20000	20000	20000	20000	20000	20000	20000	20000	20000	20000	20000	20000	20000	0.019	0.009
	0.4	1.430	0.018	0.013	0.016	0.013	0.007	0.007	0.008	0.010	0.018	0.008	0.010	0.018	0.019	0.009
	0.04	0.192	0.011	0.010	0.011	0.014	0.007	0.007	0.008	0.011	0.008	0.008	0.011	0.018	0.018	0.009
258	4	0.030	0.011	0.011	0.011	0.013	0.007	0.007	0.009	0.012	0.018	0.009	0.012	0.018	0.018	0.009
	0.4	0.042	1.831	0.114	0.020	0.015	0.009	0.007	0.010	0.012	0.019	0.010	0.012	0.019	0.019	0.010
	0.04	0.013	0.270	0.022	0.013	0.016	0.009	0.007	0.009	0.013	0.019	0.009	0.013	0.019	0.019	0.011
407	4	0.008	0.043	0.012	0.011	0.017	0.008	0.007	0.009	0.012	0.018	0.009	0.012	0.018	0.018	0.010
	0.4	0.084	0.045	0.094	0.010	0.015	0.007	0.007	0.009	0.011	0.018	0.009	0.011	0.018	0.018	0.009
	0.04	0.016	0.012	0.134	0.010	0.013	0.007	0.007	0.009	0.011	0.018	0.009	0.011	0.018	0.018	0.009
645	4	0.009	0.010	0.025	0.011	0.014	0.008	0.008	0.009	0.012	0.019	0.010	0.012	0.019	0.019	0.010
	0.4	0.149	0.034	0.023	0.032	0.016	0.008	0.008	0.009	0.012	0.020	0.009	0.012	0.020	0.020	0.010
	0.04	0.024	0.013	0.012	0.045	0.017	0.009	0.008	0.009	0.012	0.021	0.009	0.012	0.021	0.021	0.011
104	4	0.010	0.010	0.011	0.014	0.015	0.009	0.008	0.008	0.012	0.021	0.008	0.012	0.021	0.021	0.011
	0.4	0.007	0.009	0.009	0.010	0.008	0.007	0.015	0.025	0.017	0.031	0.025	0.017	0.031	0.031	0.009
	0.04	0.010	0.009	0.009	0.010	0.009	0.008	0.008	0.011	0.013	0.020	0.009	0.013	0.020	0.020	0.009
809	4	0.009	0.009	0.010	0.011	0.024	0.008	0.007	0.009	0.012	0.020	0.009	0.012	0.020	0.020	0.009
	0.4	0.013	0.012	0.012	0.011	0.114	0.877	0.047	0.143	0.046	0.080	0.143	0.046	0.080	0.080	0.017
	0.04	0.010	0.010	0.011	0.011	0.030	0.134	0.013	0.033	0.018	0.028	0.033	0.018	0.028	0.028	0.013
754	4	0.009	0.010	0.010	0.010	0.017	0.022	0.008	0.011	0.014	0.020	0.011	0.014	0.020	0.020	0.011
	0.4	0.008	0.011	0.009	0.010	0.038	0.008	0.286	0.068	0.025	0.027	0.068	0.025	0.027	0.027	0.013
	0.04	0.008	0.009	0.010	0.011	0.017	0.008	0.038	0.015	0.013	0.020	0.015	0.013	0.020	0.020	0.010
1876	4	0.009	0.009	0.011	0.011	0.016	0.008	0.011	0.010	0.012	0.020	0.010	0.012	0.020	0.020	0.009
	0.4	0.010	0.012	0.011	0.011	0.026	0.009	0.013	0.728	0.023	0.025	0.728	0.023	0.025	0.025	0.012
	0.04	0.009	0.014	0.010	0.011	0.017	0.009	0.008	0.089	0.015	0.021	0.089	0.015	0.021	0.021	0.013
47	4	0.011	0.010	0.010	0.012	0.016	0.010	0.007	0.017	0.014	0.019	0.017	0.014	0.019	0.019	0.011
	0.4	0.008	0.009	0.009	0.010	0.017	0.007	0.008	0.011	0.324	0.021	0.011	0.324	0.021	0.021	0.014
	0.04	0.008	0.009	0.009	0.011	0.015	0.008	0.008	0.009	0.048	0.020	0.009	0.048	0.020	0.020	0.013
	0.04	0.008	0.009	0.009	0.011	0.014	0.008	0.008	0.008	0.017	0.022	0.008	0.017	0.022	0.022	0.011

Table 2 (Cont'd)

Purified VLP	VLP concentration (ng/mL)	Solid-phase antibody plate x POD (top: strain name, bottom: dilution of POD-labeled antibody)											
		124 20000	258 20000	407 20000	645 20000	104 20000	809 20000	754 20000	1876 20000	47 20000	7k 20000	10-25 20000	
7k	4	0.009	0.010	0.010	0.011	0.019	0.009	0.010	0.011	0.015	0.160	0.014	
	0.4	0.009	0.011	0.010	0.011	0.016	0.008	0.008	0.008	0.015	0.035	0.016	
	0.04	0.011	0.010	0.010	0.011	0.017	0.009	0.008	0.009	0.014	0.022	0.015	
10-25	4	0.009	0.010	0.010	0.011	0.098	0.010	0.022	0.069	0.033	0.058	1.050	
	0.4	0.007	0.009	0.010	0.011	0.026	0.009	0.009	0.020	0.018	0.026	0.163	
	0.04	0.009	0.009	0.009	0.012	0.016	0.009	0.007	0.011	0.015	0.023	0.029	
Blank		0.009	0.011	0.010	0.011	0.016	0.009	0.008	0.009	0.017	0.022	0.016	

In the table, "645" indicates the Hu/NLV/Kashiwa 645/1999/JP strain, "124" the Hu/NLV/Seto 124/1989/JP strain, "258" the Hu/NLV/Funabashi 258/1996/JP strain, "407" the Hu/NLV/Chiba 407/1987/JP strain, "104" the Hu/NLV/Narita 104/1997/JP strain, "809" the Hu/NLV/Sanbu 809/1998/JP strain, "754" the Hu/NLV/Ichikawa 754/1998/JP strain, "1876" the Hu/NLV/Chitta 1876/1996/JP strain, "47" the Hu/NLV/Kashiwa 47/1997/JP strain, "7k" the Hu/NLV/Mie 7k/1994/JP strain, and "10-25" the Hu/NLV/Osaka 10-25/1999/JP strain.

As a result, no cross-reactivity was observed between viruses of the same genogroup, to say nothing of cross-reactivity between viruses of different Genogroups I and II. It was, therefore, confirmed that the serotypes of the 11 types of used virus strains were different from one another.

Test 1 Discrimination of SRSVs in Genogroup

The anti-SRSV antibodies against the SRSVs belonging to Genogroup I (the Hu/NLV/Kashiwa 645/1999/JP strain, the Hu/NLV/Seto 124/1989/JP strain, the Hu/NLV/Funabashi 258/1996/JP strain, and the Hu/NLV/Chiba 407/1987/JP strain) were diluted with a carbonate buffer (pH 9.5) to a concentration of from 0.5 to 10 μ g/mL and were then mixed. The thus-obtained mixture was poured at 100 μ L/well into a polystyrene flat-bottom microplate (manufactured by Nunc). The microplate was allowed to stand overnight at 4°C. After standing for 18 hours or longer, the microplate was washed 3 to 4 times at 200 μ L/well with PBS which contained "Tween 20" at a final concentration of 0.05%.

10 mM PBS (pH 7.2) - which contained bovine serum albumin (BSA) and "Tween 20" at final concentrations of 0.5% and 0.05%, respectively - was then added at 200 μ L/well. The microplate was allowed to stand overnight at 4°C to obtain a microplate with the anti-SRSV-IgG antibodies against the respective serotypes of Genogroup I carried in a mixed solid-phase form (Type I plate).

Next, the anti-SRSV antibodies against the SRSVs belonging to Genogroup II (the Hu/NLV/Narita 104/1997/JP strain, the Hu/NLV/Sanbu 809/1998/JP strain, the Hu/NLV/Ichikawa 754/1998/JP strain, the Hu/NLV/Chitta 1876/1996/JP strain, the Hu/NLV/Kashiwa 47/1997/JP strain, the Hu/NLV/Mie 7k/1994/JP strain, and the Hu/NLV/Osaka 10-25/1999/JP strain) were similarly formed into a solid phase to obtain a Type II plate.

To feces (0.5 to 1.0 g) of each SRSV patient, PBS (9 mL) and "Daiflon" (1 mL) were added, followed by homogenization. The thus-prepared suspension was centrifuged under 19,000 g for 20 minutes, and the supernatant was collected and formed into a 10% fecal emulsion. The 10% fecal emulsion was diluted at 1:1 in volume with a buffer. The diluted emulsion was added at 100 μ L/well into wells of the Type I and Type II plates, and was allowed to react at room temperature for 60 minutes. After the reaction, the reaction mixtures in the wells were eliminated under suction. 10 mM PBS (pH 7.2) - which contained "Tween 20" at a final concentration of 0.05% - was added at 200 μ L/well to the wells, and was then eliminated under suction. This

procedure was performed at least three times. After the washing, the POD-labeled anti-SRSV antibodies of the respective serotypes, said antibodies having had been diluted 20,000-fold with a buffer, were added at 100 μ L/well, and were then reacted at room temperature for 60 minutes. After washing, a TMB solution with hydrogen peroxide contained therein was added at 100 μ L/well, and were then reacted at room temperature for 30 minutes. Subsequent to the reaction, 0.6 N sulfuric acid was added at 100 μ L/well, and the absorbance (450 nm/630 nm) of each well was measured by an ELISA autoreader.

As a result, it was found that among 15 fecal specimens from patients infected to SRSV of Genogroup I, 14 fecal specimens reacted only to the Type I plate and did not react to the Type II plate. Concerning 7 fecal specimens from patients infected to SRSV of Genogroup II, on the other hand, 6 fecal specimens did not react to the Type I plate but reacted only to Type II plate. It has, therefore, been confirmed that discrimination in genogroup is actually feasible.

Test 2 Discrimination of SRSVs in Serotype

The anti-SRSV antibodies against the SRSVs (the Hu/NLV/Kashiwa 645/1999/JP strain, the Hu/NLV/Seto 124/1989/JP strain, the Hu/NLV/Funabashi 258/1996/JP strain, the Hu/NLV/Chiba 407/1987/JP strain, the Hu/NLV/Narita 104/1997/JP strain, the Hu/NLV/Sanbu 809/1998/JP strain, the Hu/NLV/Ichikawa 754/1998/JP strain, the Hu/NLV/Chitta 1876/1996/JP strain, the Hu/NLV/Kashiwa 47/1997/JP strain, the

Hu/NLV/Mie 7k/1994/JP strain, and the Hu/NLV/Osaka 10-25/1999/JP strain) were each independently diluted with a carbonate buffer (pH 9.5) to a concentration of from 0.5 to 10 μ g/mL. The thus-obtained dilutions were poured at 100 μ L/well into a polystyrene flat-bottom microplate (manufactured by Nunc). The microplate was allowed to stand overnight at 4°C. After having been allowed to stand for 18 hours or longer, the microplate was washed 3 to 4 times at 200 μ L/well with PBS which contained "Tween 20" at a final concentration of 0.05%. 10 mM PBS (pH 7.2) - which contained bovine serum albumin (BSA) and "Tween 20" at final concentrations of 0.5% and 0.05%, respectively - was then added at 200 μ L/well. The microplate was allowed to stand overnight at 4°C to obtain a solid-phase anti-SRSV antibody microplate (serotype discrimination plate).

With respect to fecal specimens from SRSV patients, ELISA was conducted in a similar manner as in Test 1. The results are shown in Table 3.

Table 3 Clinical Test

Total number of specimens: 41

Serotype discriminated by invention kit	Number of detected specimen(s)
HU/NLV/Kashiwa 645/1999/JP	1
Hu/NLV/Seto 124/1989/JP	7
Hu/NLV/Funabashi 258/1996/JP	4
Hu/NLV/Chiba 407/1987/JP	1
HU/NLV/Narita 104/1997/JP	4
Hu/NLV/Sanbu 809/1998/JP	12
Hu/NLV/Ichikawa 754/1998/JP	2
Hu/NLV/Chitta 1876/1996/JP	3
Hu/NLV/Kashiwa 47/1997/JP	1
Hu/NLV/Mie 7k/1994/JP	1
Hu/NLV/Osaka 10-25/1999/JP	2
Total number of detected specimens	38 (93%)

As a result, it has been found that according to the SRSV
 5 detection method of the present invention, SRSVs can be detected
 with a probability as high as 93% and their serotypes can also
 be discriminated.

Further, the serotypes discriminated by the kit of the
 present invention were consistent with those ascertained by PCR
 10 and an analysis of their base sequences (Table 4).

Table 4 Ascertainment of Serotypes

Total number of specimens: 38

Serotype discriminated by invention kit	Number of specimen(s) discriminated in serotype by PCR and analysis of base sequences
HU/NLV/Kashiwa 645/1999/JP 1 Specimens	1
Hu/NLV/Seto 124/1989/JP 7 Specimens	7
Hu/NLV/Funabashi 258/1996/JP 4 Specimens	4
Hu/NLV/Chiba 407/1987/JP 1 Specimen	1
HU/NLV/Narita 104/1997/JP 4 Specimens	4
Hu/NLV/Sanbu 809/1998/JP 12 Specimens	12
Hu/NLV/Ichikawa 754/1998/JP 2 Specimens	2
Hu/NLV/Chitta 1876/1996/JP 3 Specimens	3
Hu/NLV/Kashiwa 47/1997/JP 1 Specimen	1
Hu/NLV/Mie 7k/1994/JP 1 Specimen	1
Hu/NLV/Osaka 10-25/1999/JP 2 Specimens	2

The anti-SRSV antibodies against the SRSVs (the
 5 Hu/NLV/Kashiwa 645/1999/JP strain, the Hu/NLV/Seto 124/1989/JP
 strain, the Hu/NLV/Funabashi 258/1996/JP strain, the
 Hu/NLV/Chiba 407/1987/JP strain, the Hu/NLV/Narita 104/1997/JP
 strain, the Hu/NLV/Sanbu 809/1998/JP strain, the
 Hu/NLV/Ichikawa 754/1998/JP strain, the Hu/NLV/Chitta
 10 1876/1996/JP strain, the Hu/NLV/Kashiwa 47/1997/JP strain, the
 Hu/NLV/Mie 7k/1994/JP strain, and the Hu/NLV/Osaka
 10-25/1999/JP strain) were each independently diluted with a

carbonate buffer (pH 9.5) to a concentration of from 0.5 to 10
ig/mL. All the dilutions so obtained were mixed. As an
alternative, the anti-SRSV antibodies may be diluted after mixing
them together. Using the thus-diluted mixture of the anti-SRSV
5 antibodies, a solid-phase anti-SRSV antibody microplate was
produced likewise. With respect to 22 fecal specimens from
patients infected to SRSV, ELISA was conducted in a similar manner
as in Test 1. It was possible to detect SRSV in 20 specimens.

10 **Industrial Applicability**

According to the SRSV detection kit of the present
invention, it is possible to detect most of the SRSV-related
viruses discovered to date and also to discriminate their
serotypes and genogroups. When SRSV-related food poisoning
15 occurs, the SRSV detection kit of the present invention is,
therefore, useful for specifying an infection route, preventing
the infection from spreading, and performing an epidemiological
investigation.

CLAIMS

1. An SRSV detection kit comprising all antibodies against SRSV-related virus constituting peptides selected from the following peptide groups (a) to (k), respectively:

5 (a) a peptide having an amino acid sequence represented by SEQ ID NO: 1 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(b) a peptide having an amino acid sequence represented by SEQ ID NO: 2 and peptides each having at least 80% of homology
10 with said amino acid sequence, and partial peptides thereof,

(c) a peptide having an amino acid sequence represented by SEQ ID NO: 3 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(d) a peptide having an amino acid sequence represented
15 by SEQ ID NO: 4 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(e) a peptide having an amino acid sequence represented by SEQ ID NO: 5 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

20 (f) a peptide having an amino acid sequence represented by SEQ ID NO: 6 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(g) a peptide having an amino acid sequence represented by SEQ ID NO: 7 and peptides each having at least 80% of homology
25 with said amino acid sequence, and partial peptides thereof,

(h) a peptide having an amino acid sequence represented

by SEQ ID NO: 8 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(i) a peptide having an amino acid sequence represented by SEQ ID NO: 9 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(j) a peptide having an amino acid sequence represented by SEQ ID NO: 10 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof, and

(k) a peptide having an amino acid sequence represented by SEQ ID NO: 11 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof.

2. An SRSV detection kit according to claim 1, wherein said antibodies have been prepared by immunizing with virus-like particles.

3. An SRSV detection kit according to claim 1, which is useful for distinguishing serotype of SRSVs.

4. An SRSV detection kit for discriminating genogroup of SRSVs, the kit comprising all antibodies against SRSV-related virus constituting peptides selected from the following peptide groups (a) to (d), respectively:

(a) a peptide having an amino acid sequence represented by SEQ ID NO: 1 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(b) a peptide having an amino acid sequence represented by SEQ ID NO: 2 and peptides each having at least 80% of homology

with said amino acid sequence, and partial peptides thereof,

(c) a peptide having an amino acid sequence represented by SEQ ID NO: 3 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,
5 and

(d) a peptide having an amino acid sequence represented by SEQ ID NO: 4 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof.

5. An SRSV detection kit for discriminating genogroup
10 of SRSVs, the kit comprising all antibodies against SRSV-related virus constituting peptides selected from the following peptide groups (e) to (k), respectively:

(e) a peptide having an amino acid sequence represented by SEQ ID NO: 5 and peptides each having at least 80% of homology
15 with said amino acid sequence, and partial peptides thereof,

(f) a peptide having an amino acid sequence represented by SEQ ID NO: 6 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(g) a peptide having an amino acid sequence represented
20 by SEQ ID NO: 7 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(h) a peptide having an amino acid sequence represented by SEQ ID NO: 8 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,

(i) a peptide having an amino acid sequence represented
25 by SEQ ID NO: 9 and peptides each having at least 80% of homology

with said amino acid sequence, and partial peptides thereof,

(j) a peptide having an amino acid sequence represented by SEQ ID NO: 10 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof,
5 and

(k) a peptide having an amino acid sequence represented by SEQ ID NO: 11 and peptides each having at least 80% of homology with said amino acid sequence, and partial peptides thereof.

6. An SRSV detection kit according to claims 1-5, wherein
10 SRSVs are captured with solid-phase antibody carriers having said antibodies immobilized thereon.

7. An Hu/NLV/Chiba/407/1987/JP gene having a base sequence represented by SEQ ID NO: 15 or a base sequence similar to the first-mentioned base sequence except for deletion,
15 replacement or addition of one to several bases of said first-mentioned base sequence.

8. An Hu/NLV/Kashiwa 47/1997/JP gene having a base sequence represented by SEQ ID NO: 20 or a base sequence similar to the first-mentioned base sequence except for deletion,
20 replacement or addition of one to several bases of said first-mentioned base sequence.

9. An Hu/NLV/Mie 7k/1994/JP gene having a base sequence represented by SEQ ID NO: 21 or a base sequence similar to the first-mentioned base sequence except for deletion, replacement
25 or addition of one to several bases of said first-mentioned base sequence.

10. An Hu/NLV/Osaka 10-25/1999/JP gene having a base sequence represented by SEQ ID NO: 22 or a base sequence similar to the first-mentioned base sequence except for deletion, replacement or addition of one to several bases of said
5 first-mentioned base sequence.

10-25/1999/JP

ABSTRACT

This invention relates to an SRSV detection kit comprising all antibodies against SRSV-related virus constituting peptides selected from the following peptide groups (a) to (k),

5 respectively: (a) a peptide having an amino acid sequence represented by SEQ ID NO: 1, and the like, (b) a peptide having an amino acid sequence represented by SEQ ID NO: 2, and the like, (c) a peptide having an amino acid sequence represented by SEQ ID NO: 3, and the like, (d) a peptide having an amino acid sequence represented by SEQ ID NO: 4, and the like, (e) a peptide having an amino acid sequence represented by SEQ ID NO: 5, and the like, (f) a peptide having an amino acid sequence represented by SEQ ID NO: 6, and the like, (g) a peptide having an amino acid sequence represented by SEQ ID NO: 7, and the like, (h) a peptide having an amino acid sequence represented by SEQ ID NO: 8, and the like, (i) a peptide having an amino acid sequence represented by SEQ ID NO: 9, and the like, (j) a peptide having an amino acid sequence represented by SEQ ID NO: 10, and the like, and (k) a peptide having an amino acid sequence represented by SEQ ID NO: 11, and the like.

Use of the kit makes it possible to detect most SRSV-related viruses and further to distinguish their serotypes and genogroups, easily and surely.

FIG. 1

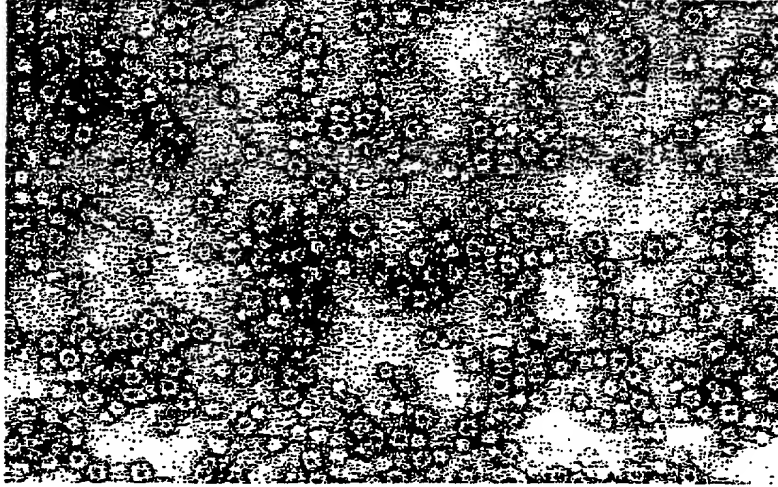


FIG. 2

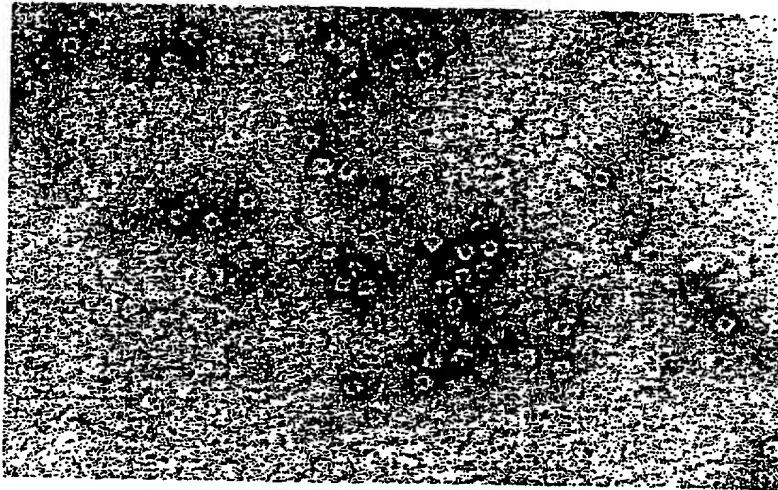


FIG. 3

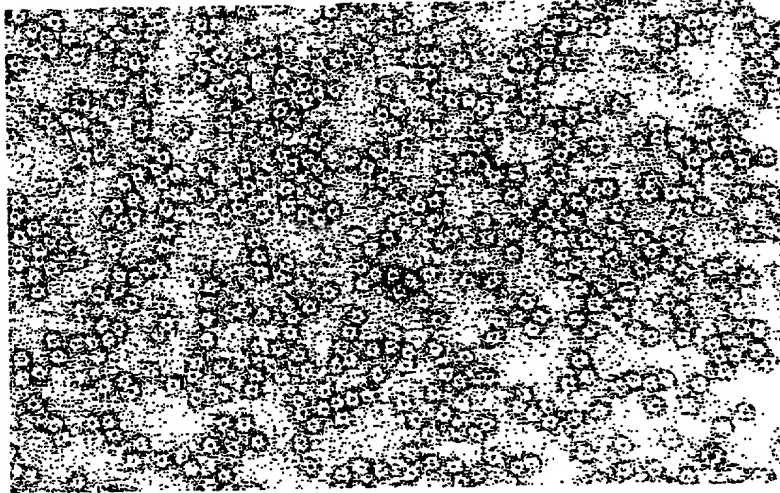


FIG. 4

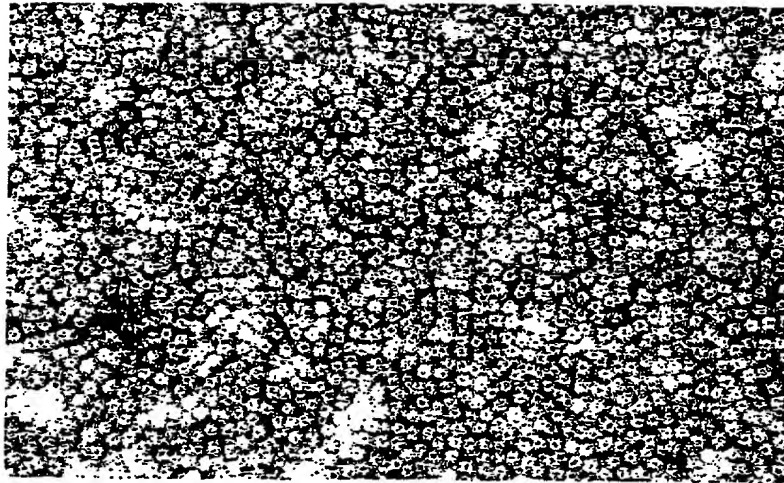


FIG. 5

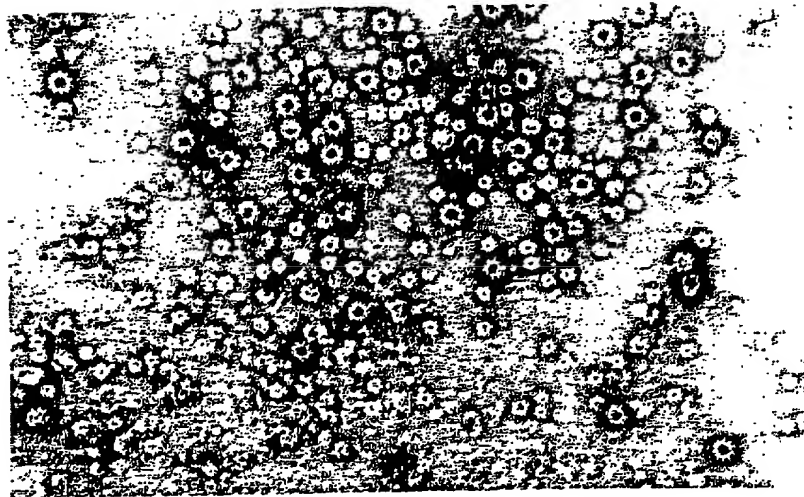


FIG. 6

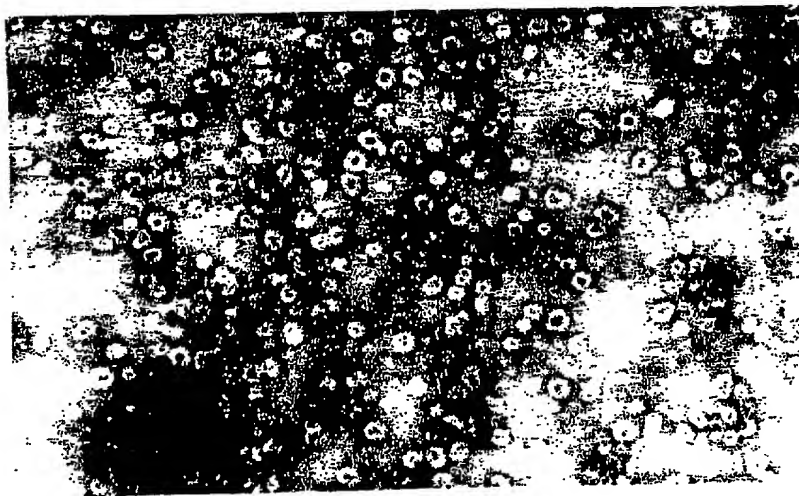


FIG. 7

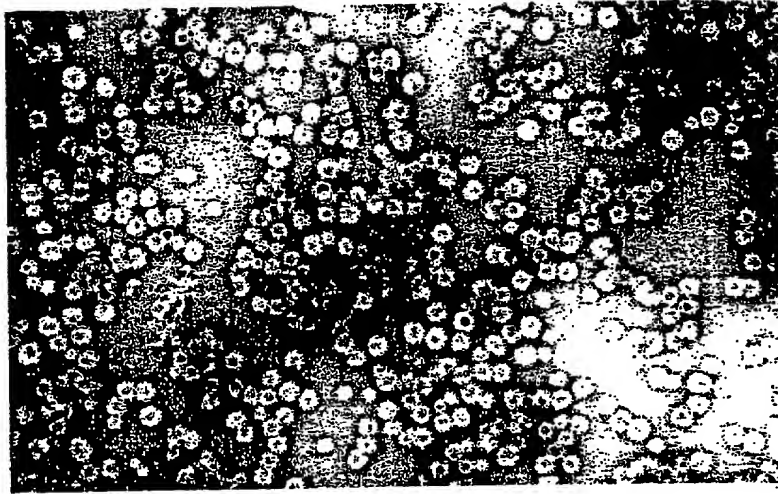


FIG. 8

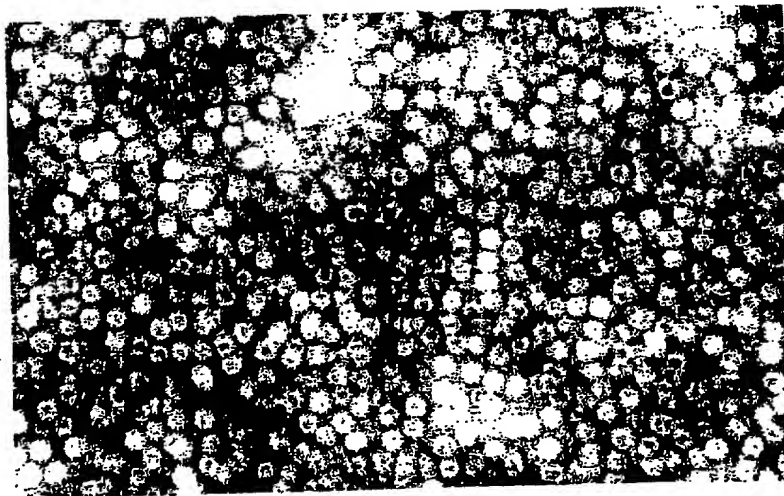


FIG. 9

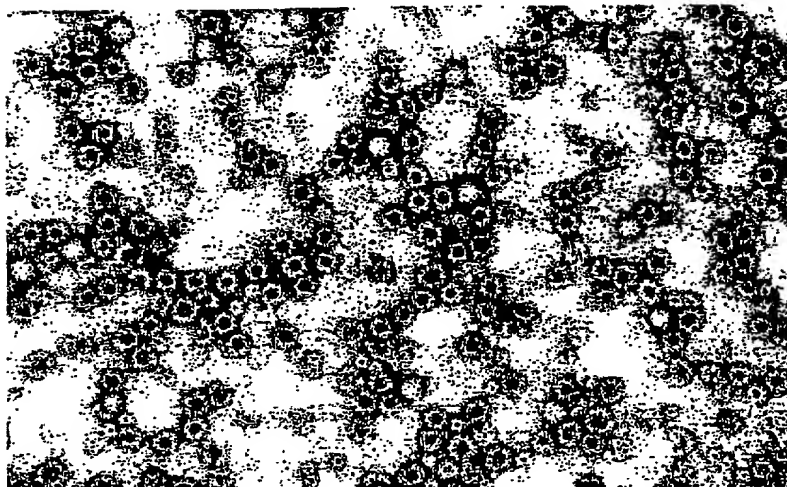
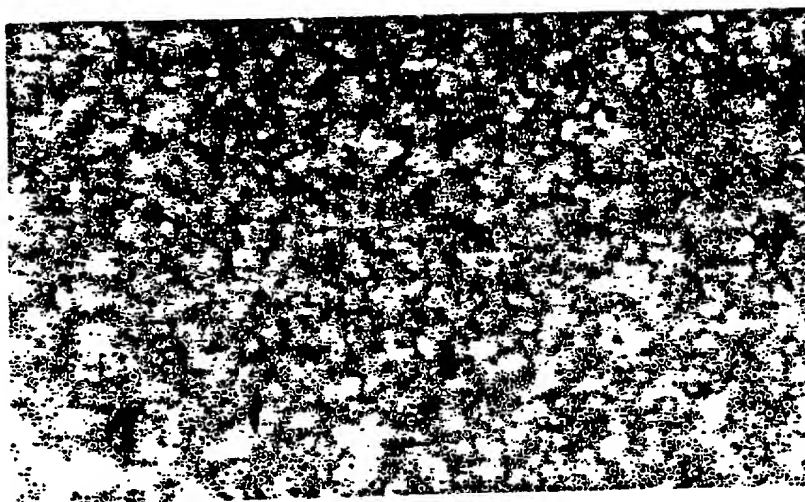


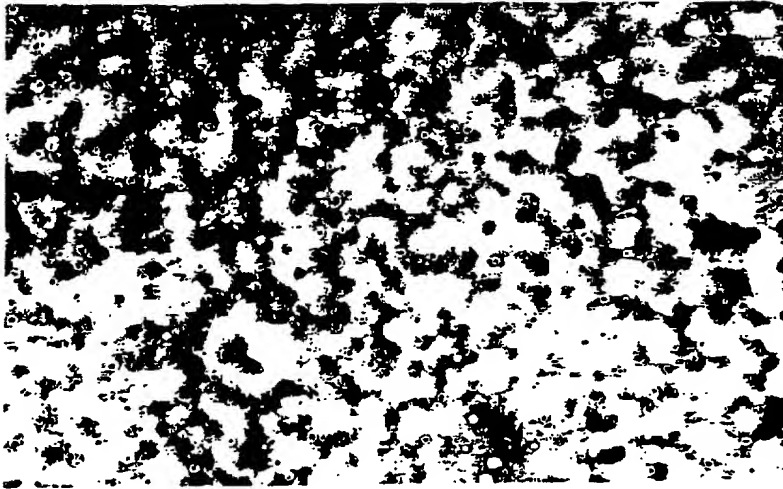
FIG. 10



09926799-122001

09/926799

FIG. 11



Declaration and Power of Attorney For Patent Application

特許出願宣言書及び委任状

DK0001 USA ①

Japanese Language Declaration

日本語宣言書

下記の氏名の発明者として、私は以下の通り宣言します。

As a below named inventor, I hereby declare that:

私の住所、私書箱、国籍は下記の私の氏名の後に記載された通りです。

My residence, post office address and citizenship are as stated next to my name.

下記の名称の発明に関して請求範囲に記載され、特許出願している発明内容について、私が最初かつ唯一の発明者（下記の氏名が一つの場合）もしくは最初かつ共同発明者（下記の名称が複数の場合）であると信じています。

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled.

S R S V 検出キット

SRSV DETECTION KIT

上記発明の明細書は、

the specification of which

☐ 本書に添付されています。

☐ is attached hereto.

☒ 2000 6 月 22 日に提出され、米国出願番号または特許協定条約国際出願番号を PCT/JP00/04095 とし、

☒ was filed on June 22, 2000

(該当する場合) _____ に訂正されました。

as ~~United States Application Number~~

PCT International Application Number

PCT/JP00/04095

and was amended on

(if applicable).

私は、特許請求範囲を含む上記訂正後の明細書を検討し、内容を理解していることをここに表明します。

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

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I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56.

Japanese Language Declaration
(日本語宣言書)

私は、米国法典第35編119条 (a) - (d) 項又は365条 (b) 項に基づき下記の、米国以外の国の少なくとも一カ国を指定している特許協力条約365 (a) 項に基づく国際出願、又は外国での特許出願もしくは発明者証の出願についての外国優先権をここに主張するとともに、優先権を主張している、本出願の前に出願された特許または発明者証の外国出願を以下に、枠内をマークすることで、示しています。

Prior Foreign Application(s)

外国での先行出願

11-175928

(Number)
(番号)

Japan

(Country)
(国名)

(Number)
(番号)

(Country)
(国名)

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(Application No.)
(出願番号)

(Filing Date)
(出願日)

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(Application No.)
(出願番号)

(Filing Date)
(出願日)

(Application No.)
(出願番号)

(Filing Date)
(出願日)

私は、私自信の知識に基づいて本宣言書中で私が行なう表明が真実であり、かつ私の入手した情報と私の信じるところに基づく表明が全て真実であると信じていること、さらに故意になされた虚偽の表明及びそれと同等の行為は米国法典第18編第1001条に基づき、罰金または拘禁、もしくはその両方により処罰されること、そしてそのような故意による虚偽の声明を行なえば、出願した、又は既に許可された特許の有効性が失われることを認識し、よってここに上記のごとく宣誓を致します。

I hereby claim foreign priority under Title 35, United States Code, Section 119 (a)-(d) or 365(b) of any foreign application(s) for patent or inventor's certificate, or Section 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed.

Priority Claimed

優先権主張

☒

☐

Yes

No

はい

いいえ

☐

☐

Yes

No

はい

いいえ

22/06/1999

(Day/Month/Year Filed)
(出願年月日)

(Day/Month/Year Filed)
(出願年月日)

I hereby claim the benefit under Title 35, United States Code, Section 119(e) of any United States provisional application(s) listed below.

(Application No.)
(出願番号)

(Filing Date)
(出願日)

I hereby claim the benefit under Title 35, United States Code, Section 120 of any United States application(s), or Section 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of Title 35, United States Code Section 112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of application.

(Status: Patented, Pending, Abandoned)
(現況: 特許許可済、係属中、放棄済)

(Status: Patented, Pending, Abandoned)
(現況: 特許許可済、係属中、放棄済)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Japanese Language Declaration

(日本語宣言書)

委任状：私は下記の発明者として、本出願に関する一切の手続きを米特許商標局に対して遂行する弁理士または代理人として、下記の者を指名いたします。
(弁護士、または代理人の指名及び登録番号を明記のこと)

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith: (list name and registration number)

**022850**

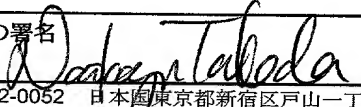
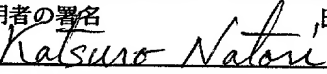
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Send Correspondence to:

**022850**

直接電話連絡先：(名前及び電話番号)

Direct Telephone Calls to: (name and telephone number)
(703) 413-3000

単独発明者または第一の共同発明者の氏名 武田 直和 1-00		Full name of sole or first joint inventor Naokazu TAKEDA	
発明者の署名  日付 Nov. 9, 2001		Inventor's signature Date	
住所 162-0052 日本国東京都新宿区戸山一丁目 23 番 1 号 国立感染症研究所内		Residence C/O NATIONAL INSTITUTE OF INFECTIOUS DISEASES 23-1, TOYAMA 1-CHOME, SHINJUKU-KU, TOKYO JPX 162-0052 JAPAN	
国籍 日本国		Citizenship JAPANESE	
郵便の宛先 住所に同じ		Post Office Address SAME AS ABOVE	
第二の共同発明者の氏名 名取 克郎 2-00		Full name of second joint inventor, if any Katsuro NATORI	
第二の共同発明者の署名  日付 Nov. 9, 2001		Second joint Inventor's signature Date	
住所 162-0052 日本国東京都新宿区戸山一丁目 23 番 1 号 国立感染症研究所内		Residence C/O NATIONAL INSTITUTE OF INFECTIOUS DISEASES 23-1, TOYAMA 1-CHOME, SHINJUKU-KU, TOKYO 162-0052 JAPAN	
国籍 日本国		Citizenship JAPANESE JPX	
郵便の宛先 住所に同じ		Post Office Address SAME AS ABOVE	

(第三以降の共同発明者についても同様に記載し、署名すること)

(Supply similar information and signature for third and subsequent joint inventors.)

Japanese Language Declaration

(日本語宣言書)

第三の共同発明者の氏名	宮村 達男 300	Full name of third joint inventor, if any	Tatsuo MIYAMURA
第三の共同発明者の署名	日付 Nov. 9, 2001	Third joint Inventor's signature	Date
住所 162-0052 日本国東京都新宿区戸山一丁目23番1号 国立感染症研究所内		Residence	C/O NATIONAL INSTITUTE OF INFECTIOUS DISEASES 23-1, TOYAMA 1-CHOME, SHINJUKU-KU, TOKYO
国籍 日本国		Citizenship	162-0052 JAPAN JAPANESE JPK
郵便の宛先 住所に同じ		Post Office Address	SAME AS ABOVE

第四の共同発明者の氏名	鎌田 公仁夫 400	Full name of fourth joint inventor, if any	Kunio KAMATA
第四の共同発明者の署名	日付 Oct. 29, 2001	Fourth joint Inventor's signature	Date
住所 959-1836 日本国新潟県五泉市南本町一丁目2番2号 デンカ生研株式会社内		Residence	C/O DENKA SEIKEN CO., LTD., 2-2, MINAMIHONCHO 1-CHOME, GOSEN-SHI, NIIGATA 959-1836 JAPAN
国籍 日本国		Citizenship	JAPANESE JPK
郵便の宛先 住所に同じ		Post Office Address	SAME AS ABOVE

第五の共同発明者の氏名	佐藤 俊則 500	Full name of fifth joint inventor, if any	Toshinori SATO
第五の共同発明者の署名	日付 Oct. 31, 2001	Fifth joint Inventor's signature	Date
住所 959-1836 日本国新潟県五泉市南本町一丁目2番2号 デンカ生研株式会社内		Residence	C/O DENKA SEIKEN CO., LTD., 2-2, MINAMIHONCHO 1-CHOME, GOSEN-SHI, NIIGATA 959-1836 JAPAN
国籍 日本国		Citizenship	JAPANESE JPK
郵便の宛先 住所に同じ		Post Office Address	SAME AS ABOVE

第六の共同発明者の氏名	佐藤 征也 600	Full name of sixth joint inventor, if any	Seiya SATO
第六の共同発明者の署名	日付 Oct 31 2001	Sixth joint Inventor's signature	Date
住所 959-1836 日本国新潟県五泉市南本町一丁目2番2号 デンカ生研株式会社内		Residence	C/O DENKA SEIKEN CO., LTD., 2-2, MINAMIHONCHO 1-CHOME, GOSEN-SHI, NIIGATA 959-1836 JAPAN
国籍 日本国		Citizenship	JAPANESE JPK
郵便の宛先 住所に同じ		Post Office Address	SAME AS ABOVE

(第六またはそれ以降の共同発明者に対しても同様な情報および署名を提供すること。)

(Supply similar information and signature for third and subsequent joint inventors.)

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Ala Phe Thr Ala Gly Lys Ile Ile Phe Ala Ala Ile Pro Pro Asn Phe

115	120	125
Pro Ile Asp Asn Leu Ser Ala Ala Gln Ile Thr Met Cys Pro His Val		
130	135	140
Ile Val Asp Val Arg Gln Leu Glu Pro Val Asn Leu Pro Met Pro Asp		
145	150	155 160
Val Arg Asn Asn Phe Phe His Tyr Asn Gln Gly Ser Asp Ser Arg Leu		
	165 170	175
Arg Leu Ile Ala Met Leu Tyr Thr Pro Leu Arg Ala Asn Asn Ser Gly		
	180 185	190
Asp Asp Val Phe Thr Val Ser Cys Arg Val Leu Thr Arg Pro Ser Pro		
	195 200	205
Asp Phe Ser Phe Asn Phe Leu Val Pro Pro Thr Val Glu Ser Lys Thr		
	210 215	220
Lys Pro Phe Thr Leu Pro Ile Leu Thr Ile Ser Glu Met Ser Asn Ser		
	225 230	235 240
Arg Phe Pro Val Pro Ile Glu Ser Leu His Thr Ser Pro Thr Glu Asn		
	245 250	255
Ile Val Val Gln Cys Gln Asn Gly Arg Val Thr Leu Asp Gly Glu Leu		
	260 265	270
Met Gly Thr Thr Gln Leu Leu Pro Ser Gln Ile Cys Ala Phe Arg Gly		
	275 280	285
Val Leu Thr Arg Ser Thr Ser Arg Ala Ser Asp Gln Ala Asp Thr Ala		
	290 295	300
Thr Pro Arg Leu Phe Asn Tyr Tyr Trp His Val Gln Leu Asp Asn Leu		
	305 310	315 320
Asn Gly Thr Pro Tyr Asp Pro Ala Glu Asp Ile Pro Gly Pro Leu Gly		
	325 330	335
Thr Pro Asp Phe Arg Gly Lys Val Phe Gly Val Ala Ser Gln Arg Asn		
	340 345	350
Leu Asp Ser Thr Thr Arg Ala His Glu Ala Lys Val Asp Thr Thr Ala		
	355 360	365

Gly Arg Phe Thr Pro Lys Leu Gly Ser Leu Glu Ile Ser Thr Asp Ser
370 375 380

Asp Asp Phe Asp Gln Asn Gln Pro Thr Lys Phe Thr Pro Val Gly Ile
385 390 395 400

Gly Val Asp Asn Glu Ala Glu Phe Gln Gln Trp Ser Leu Pro Asp Tyr
405 410 415

Ser Gly Gln Phe Thr His Asn Met Asn Leu Ala Pro Ala Val Ala Pro
420 425 430

Asn Phe Pro Gly Glu Gln Leu Leu Phe Phe Arg Ser Gln Leu Pro Ser
435 440 445

Ser Gly Gly Arg Ser Asn Gly Val Leu Asp Cys Leu Val Pro Gln Glu
450 455 460

Trp Val Gln His Phe Tyr Gln Glu Ser Ala Pro Ala Gln Thr Gln Val
465 470 475 480

Ala Leu Val Arg Tyr Val Asn Pro Asp Thr Gly Lys Val Leu Phe Glu
485 490 495

Ala Lys Leu His Lys Leu Gly Phe Met Thr Ile Ala Asn Asn Gly Asp
500 505 510

Ser Pro Ile Thr Val Pro Pro Asn Gly Tyr Phe Arg Phe Glu Ser Trp
515 520 525

Val Asn Pro Phe Tyr Thr Leu Ala Pro Met Gly Thr Gly Asn Gly Arg
530 535 540

Arg Arg Ile Gln
545

<210> 7

<211> 540

<212> PRT

<213> Hu/NLV/Ichikawa 754/1998/JP

<400> 7

Met Lys Met Ala Ser Asn Asp Ala Thr Pro Ser Asn Asp Gly Ala Ala
1 5 10 15

Gly Leu Val Pro Glu Ser Asn Asn Glu Ala Met Ala Leu Glu Pro Val
 20 25 30

Val Gly Ala Ser Leu Ala Ala Pro Val Thr Gly Gln Thr Asn Ile Ile
 35 40 45

Asp Pro Trp Ile Arg Thr Asn Phe Val Gln Ala Pro Asn Gly Glu Phe
 50 55 60

Thr Val Ser Pro Arg Asn Ser Pro Gly Glu Ile Leu Val Asn Leu Glu
 65 70 75 80

Leu Gly Pro Glu Leu Asn Pro Tyr Leu Ala His Leu Ala Arg Met Tyr
 85 90 95

Asn Gly Tyr Ala Gly Gly Met Glu Val Gln Val Met Leu Ala Gly Asn
 100 105 110

Ala Phe Thr Ala Gly Lys Ile Ile Phe Ala Ala Val Pro Pro Tyr Phe
 115 120 125

Pro Val Glu Asn Leu Ser Pro Ser Gln Ile Thr Met Phe Pro His Val
 130 135 140

Ile Ile Asp Val Arg Thr Leu Glu Pro Val Leu Leu Pro Met Pro Asp
 145 150 155 160

Val Arg Ser Thr Leu Phe His Phe Asn Gln Lys Asp Glu Pro Lys Met
 165 170 175

Arg Leu Val Ala Met Leu Tyr Thr Pro Leu Arg Ser Asn Gly Ser Gly
 180 185 190

Asp Asp Val Phe Thr Val Ser Cys Arg Ile Leu Thr Arg Pro Ser Pro
 195 200 205

Glu Phe Asp Phe Thr Tyr Leu Val Pro Pro Thr Val Glu Ser Lys Thr
 210 215 220

Lys Pro Phe Thr Leu Pro Val Leu Thr Leu Gly Glu Leu Ser Asn Ser
 225 230 235 240

Arg Phe Pro Leu Ser Ile Asp Glu Met Val Thr Ser Pro Asn Glu Ser
 245 250 255

Ile Val Val Gln Pro Gln Asn Gly Arg Val Thr Leu Asp Gly Glu Leu
260 265 270

Leu Gly Thr Thr Gln Leu Gln Ala Cys Asn Ile Cys Ser Ile Arg Gly
275 280 285

Lys Val Thr Gly Gln Val Pro Ser Glu Gln His Met Trp Asn Leu Glu
290 295 300

Ile Thr Asn Leu Asn Gly Thr Gln Phe Asp Pro Thr Asp Asp Val Pro
305 310 315 320

Ala Pro Leu Gly Val Pro Asp Phe Ala Gly Glu Val Phe Gly Val Leu
325 330 335

Ser Gln Arg Asn Arg Gly Glu Ser Asn Pro Ala Asn Arg Ala His Asp
340 345 350

Ala Val Val Ala Thr Tyr Ser Asp Lys Tyr Thr Pro Lys Leu Gly Leu
355 360 365

Val Gln Ile Gly Thr Trp Asn Thr Asn Asp Val Glu Asn Gln Pro Thr
370 375 380

Lys Phe Thr Pro Ile Gly Leu Asn Glu Val Ala Asn Gly His Arg Phe
385 390 395 400

Glu Gln Trp Thr Leu Pro Arg Tyr Ser Gly Ala Leu Thr Leu Asn Met
405 410 415

Asn Leu Ala Pro Ala Val Ala Pro Leu Phe Pro Gly Glu Arg Leu Leu
420 425 430

Phe Phe Arg Ser Tyr Val Pro Leu Lys Gly Gly Phe Gly Asn Pro Ala
435 440 445

Ile Asp Cys Ser Val Pro Gln Glu Trp Val Gln His Phe Tyr Gln Glu
450 455 460

Ser Ala Pro Ser Leu Gly Asp Val Ala Leu Val Arg Tyr Val Asn Pro
465 470 475 480

Asp Thr Gly Arg Val Leu Phe Glu Ala Lys Leu His Lys Gly Gly Phe
485 490 495

Leu Thr Val Ser Ser Thr Ser Thr Gly Pro Val Val Val Pro Ala Asn

500

505

510

Gly Tyr Phe Lys Phe Asp Ser Trp Val Asn Gln Phe Tyr Ser Leu Ala
515 520 525

Pro Met Gly Thr Gly Asn Gly Arg Arg Arg Val Gln
530 535 540

<210> 8

<211> 535

<212> PRT

<213> Hu/NLV/Chitta 1876/1996/JP

<400> 8

Met Lys Met Ala Ser Asn Asp Ala Ala Pro Ser Asn Asp Gly Ala Ala
1 5 10 15

Gly Leu Val Pro Glu Ala Asn Asn Glu Thr Met Ala Leu Glu Pro Val
20 25 30

Ala Gly Ala Ser Ile Ala Ala Pro Leu Thr Gly Gln Asn Asn Ile Ile
35 40 45

Asp Pro Trp Ile Arg Leu Asn Phe Val Gln Ala Pro Asn Gly Glu Phe
50 55 60

Thr Val Ser Pro Arg Asn Ser Pro Gly Glu Val Leu Leu Asn Leu Glu
65 70 75 80

Leu Gly Pro Glu Leu Asn Pro Tyr Leu Ala His Leu Ser Arg Met Tyr
85 90 95

Asn Gly Tyr Ala Gly Gly Val Glu Val Gln Val Leu Leu Ala Gly Asn
100 105 110

Ala Phe Thr Ala Gly Lys Leu Val Phe Ala Ala Val Pro Pro His Phe
115 120 125

Pro Leu Glu Asn Ile Ser Pro Gly Gln Ile Thr Met Phe Pro His Val
130 135 140

Ile Ile Asp Val Arg Thr Leu Glu Pro Val Leu Leu Pro Leu Pro Asp
145 150 155 160

Val Arg Asn Asn Phe Phe His Tyr Asn Gln Gln Asn Glu Pro Arg Met

165	170	175
Arg Leu Val Ala Met Leu Tyr Thr Pro Leu Arg Ser Asn Gly Ser Gly		
180	185	190
Asp Asp Val Phe Thr Val Ser Cys Arg Val Leu Thr Arg Pro Ser Pro		
195	200	205
Asp Phe Asp Phe Asn Tyr Leu Val Pro Pro Thr Leu Glu Ser Lys Thr		
210	215	220
Lys Pro Phe Thr Leu Pro Ile Leu Thr Ile Gly Glu Leu Thr Asn Ser		
225	230	235
Arg Phe Pro Val Pro Ile Asp Glu Leu Tyr Thr Ser Pro Asn Glu Ser		
245	250	255
Leu Val Val Gln Pro Gln Asn Gly Arg Cys Ala Leu Asp Gly Glu Leu		
260	265	270
Gln Gly Thr Thr Gln Leu Leu Pro Thr Ala Ile Cys Ser Phe Arg Gly		
275	280	285
Arg Ile Asn Gln Lys Val Ser Gly Glu Asn His Val Trp Asn Met Gln		
290	295	300
Val Thr Asn Ile Asn Gly Thr Pro Phe Asp Pro Thr Gly Asp Val Pro		
305	310	315
Ala Pro Leu Gly Thr Pro Asp Phe Ser Gly Lys Leu Phe Gly Val Leu		
325	330	335
Ser Gln Arg Asp His Asp Asn Ala Cys Arg Ser His Asp Ala Val Ile		
340	345	350
Ala Thr Asn Ser Ala Lys Phe Thr Pro Lys Leu Gly Ala Ile Gln Ile		
355	360	365
Gly Thr Trp Glu Glu Asp Asp Val His Ile Asn Gln Pro Thr Lys Phe		
370	375	380
Thr Pro Val Gly Leu Phe Glu Asn Glu Gly Phe Asn Gln Trp Thr Leu		
385	390	395
Pro Asn Tyr Ser Gly Ala Leu Thr Leu Asn Met Gly Leu Ala Pro Pro		
405	410	415

Val Ala Pro Thr Phe Pro Gly Glu Gln Ile Leu Phe Phe Arg Ser His
420 425 430

Ile Pro Leu Lys Gly Gly Val Ala Asp Pro Val Ile Asp Cys Leu Leu
435 440 445

Pro Gln Glu Trp Ile Gln His Leu Tyr Gln Glu Ser Ala Pro Ser Gln
450 455 460

Ser Asp Val Ala Leu Ile Arg Phe Thr Asn Pro Asp Thr Gly Arg Val
465 470 475 480

Leu Phe Glu Ala Lys Leu His Arg Ser Gly Tyr Ile Thr Val Ala Asn
485 490 495

Thr Gly Ser Arg Pro Ile Val Val Pro Ala Asn Gly Tyr Phe Arg Phe
500 505 510

Asp Thr Trp Val Asn Gln Phe Tyr Ser Leu Ala Pro Met Gly Thr Gly
515 520 525

Asn Gly Arg Arg Arg Val Gln
530 535

<210> 9

<211> 542

<212> PRT

<213> Hu/NLV/Kashiwa 47/1997/JP

<400> 9

Met Lys Met Ala Ser Asn Asp Ala Ala Pro Ser Asn Asp Gly Ala Ala
1 5 10 15

Ser Leu Val Pro Glu Gly Ile Asn Glu Thr Met Pro Leu Glu Pro Val
20 25 30

Ala Gly Ala Ser Ile Ala Ala Pro Val Ala Gly Gln Thr Asn Ile Ile
35 40 45

Asp Pro Trp Ile Arg Thr Asn Phe Val Gln Ala Pro Asn Gly Glu Phe
50 55 60

Thr Val Ser Pro Arg Asn Ser Pro Gly Glu Ile Leu Leu Asn Leu Glu
65 70 75 80

Leu Gly Pro Asp Leu Asn Pro Tyr Leu Ala His Leu Ser Arg Met Tyr
85 90 95

Asn Gly Tyr Ala Gly Gly Val Glu Val Gln Val Leu Leu Ala Gly Asn
100 105 110

Ala Phe Thr Ala Gly Lys Ile Leu Phe Ala Ala Ile Pro Pro Asn Phe
115 120 125

Leu Val Asp Met Ile Ser Pro Ala Gln Ile Thr Met Leu Pro His Leu
130 135 140

Ile Val Asp Val Arg Thr Leu Glu Pro Ile Met Thr Pro Leu Pro Asp
145 150 155 160

Val Arg Asn Val Phe Tyr His Phe Asn Asn Gln Pro Gln Pro Arg Met
165 170 175

Arg Leu Val Ala Met Leu Tyr Thr Pro Leu Arg Ser Asn Gly Ser Gly
180 185 190

Asp Asp Val Phe Thr Val Ser Cys Arg Val Leu Thr Arg Pro Thr Pro
195 200 205

Asp Phe Glu Phe Ile Tyr Leu Val Pro Pro Ser Val Glu Ser Lys Thr
210 215 220

Lys Pro Phe Thr Leu Pro Ile Leu Thr Ile Ser Glu Leu Thr Asn Ser
225 230 235 240

Arg Phe Pro Ile Pro Ile Glu Gln Leu Tyr Thr Ala Pro Asn Glu Thr
245 250 255

Asn Val Val Gln Cys Gln Asn Gly Arg Cys Thr Leu Asp Gly Glu Leu
260 265 270

Gln Gly Thr Thr Gln Leu Leu Ser Ser Ala Val Cys Phe Leu Gln Gly
275 280 285

Arg Thr Val Ala Asp Asn Gly Asp Asn Trp Asp Gln Asn Leu Leu Gln
290 295 300

Leu Thr Tyr Pro Asn Gly Ala Ser Tyr Asp Pro Thr Asp Glu Val Pro
305 310 315 320

<213> Hu/NLV/Mie 7k/1994/JP

<400> 10

Met Lys Met Ala Ser Asn Asp Ala Ala Pro Ser Asn Asp Gly Ala Ala
1 5 10 15

Asn Leu Val Pro Glu Ala Asn Asp Glu Val Met Ala Leu Glu Pro Val
20 25 30

Val Gly Ala Ser Ile Ala Ala Pro Val Val Gly Gln Gln Asn Ile Ile
35 40 45

Asp Pro Trp Ile Arg Glu Asn Phe Val Gln Ala Pro Gln Gly Glu Phe
50 55 60

Thr Val Ser Pro Arg Asn Ser Pro Gly Glu Met Leu Leu Asn Leu Glu
65 70 75 80

Leu Gly Pro Glu Leu Asn Pro Tyr Leu Ser His Leu Ser Arg Met Tyr
85 90 95

Asn Gly Tyr Ala Gly Gly Met Gln Val Gln Val Val Leu Ala Gly Asn
100 105 110

Ala Phe Thr Ala Gly Lys Ile Ile Phe Ala Ala Val Pro Pro His Phe
115 120 125

Pro Val Glu Asn Ile Ser Ala Ala Gln Ile Thr Met Cys Pro His Val
130 135 140

Ile Val Asp Val Arg Gln Leu Glu Pro Val Leu Leu Pro Leu Pro Asp
145 150 155 160

Ile Arg Asn Arg Phe Phe His Tyr Asn Gln Glu Asn Thr Pro Arg Met
165 170 175

Arg Leu Val Ala Met Leu Tyr Thr Pro Leu Arg Ala Asn Ser Gly Glu
180 185 190

Asp Val Phe Thr Val Ser Cys Arg Val Leu Thr Arg Pro Ala Pro Asp
195 200 205

Phe Glu Phe Thr Phe Leu Val Pro Pro Thr Val Glu Ser Lys Thr Lys
210 215 220

Pro Phe Thr Leu Pro Ile Leu Thr Leu Gly Glu Leu Ser Asn Ser Arg

66292550

225	230	235	240
Phe Pro Ala Ala Ile Asp Met Leu Tyr Thr Asp Pro Asn Glu Ser Ile	245	250	255
Val Val Gln Pro Gln Asn Gly Arg Cys Thr Leu Asp Gly Thr Leu Gln	260	265	270
Gly Thr Thr Gln Leu Val Pro Thr Gln Ile Cys Ala Phe Arg Gly Thr	275	280	285
Leu Ile Ser Gln Thr Ala Arg Ala Ala Asp Ser Thr Asp Ser Pro Gln	290	295	300
Arg Ala Arg Asn His Pro Leu His Val Gln Val Lys Asn Leu Asp Gly	305	310	315
Thr Gln Tyr Asp Pro Thr Asp Asp Ile Pro Ala Val Leu Gly Ala Ile	325	330	335
Asp Phe Lys Gly Thr Val Phe Gly Val Ala Ser Gln Arg Asp Val Ser	340	345	350
Gly Gln Gln Glu Gln Gly His Tyr Ala Thr Arg Ala His Glu Ala His	355	360	365
Ile Asp Thr Thr Asp Pro Lys Tyr Ala Pro Lys Leu Gly Thr Ile Leu	370	375	380
Ile Lys Ser Gly Ser Asp Asp Phe Asn Thr Asn Gln Pro Ile Arg Phe	385	390	395
Thr Pro Val Gly Met Gly Asp Asn Asn Trp Arg Gln Trp Glu Leu Pro	405	410	415
Asp Tyr Ser Gly Arg Leu Thr Leu Asn Met Asn Leu Ala Pro Ala Val	420	425	430
Ser Pro Ser Phe Pro Gly Glu Arg Ile Leu Phe Phe Arg Ser Ile Val	435	440	445
Pro Ser Ala Gly Gly Tyr Gly Ser Gly Tyr Ile Asp Cys Leu Ile Pro	450	455	460
Gln Glu Trp Val Gln His Phe Tyr Gln Glu Ala Ala Pro Ser Gln Ser	465	470	475
			480

Ala Val Ala Leu Val Arg Tyr Val Asn Pro Asp Thr Gly Arg Asn Ile
485 490 495

Phe Glu Ala Lys Leu His Arg Glu Gly Phe Leu Thr Val Ala Asn Cys
500 505 510

Gly Asn Asn Pro Ile Val Val Pro Pro Asn Gly Tyr Phe Arg Phe Glu
515 520 525

Ala Trp Gly Asn Gln Phe Tyr Thr Leu Ala Pro Met Gly Ser Gly Gln
530 535 540

Gly Arg Arg Arg Ala Gln
545 550

<210> 11

<211> 541

<212> PRT

<213> Hu/NLV/Osaka 10-25/1999/JP

<400> 11

Met Lys Met Ala Ser Asn Asp Ala Ala Pro Ser Ser Asp Gly Ala Ala
1 5 10 15

Gly Leu Val Pro Glu Ile Asn Asn Glu Val Met Pro Leu Glu Pro Val
20 25 30

Ala Gly Ala Ser Leu Ala Thr Pro Val Val Gly Gln Gln Asn Ile Ile
35 40 45

Asp Pro Trp Ile Arg Asn Asn Phe Val Gln Ala Pro Ala Gly Glu Phe
50 55 60

Thr Val Ser Pro Arg Asn Ser Pro Gly Glu Ile Leu Leu Asp Leu Glu
65 70 75 80

Leu Gly Pro Asp Leu Asn Pro Tyr Leu Ala His Leu Ala Arg Met Tyr
85 90 95

Asn Gly His Ala Gly Gly Met Glu Val Gln Ile Val Leu Ala Gly Asn
100 105 110

Ala Phe Thr Ala Gly Lys Ile Ile Phe Ala Ala Ile Pro Pro Gly Phe
115 120 125

Pro Tyr Glu Asn Leu Ser Pro Ser Gln Ile Thr Met Cys Pro His Val	130	135	140
Ile Ile Asp Val Arg Gln Leu Glu Pro Phe Leu Leu Pro Met Pro Asp	145	150	155 160
Ile Trp Asn Asn Phe Phe His Tyr Asn Gln Gly Asn Asp Pro Lys Leu	165	170	175
Arg Leu Val Ala Met Leu Tyr Thr Pro Leu Arg Ala Asn Asn Ser Gly	180	185	190
Asp Asp Val Phe Thr Val Ser Cys Arg Val Leu Thr Lys Pro Ser Pro	195	200	205
Asp Phe Glu Phe Thr Phe Leu Val Pro Pro Thr Val Glu Ser Lys Thr	210	215	220
Lys Gln Phe Ala Leu Pro Ile Leu Lys Ile Ser Glu Met Thr Asn Ser	225	230	235 240
Arg Phe Pro Val Pro Val Asp Val Met Tyr Thr Ala Arg Asn Glu Asn	245	250	255
Gln Val Val Gln Pro Gln Asn Gly Arg Val Thr Leu Asp Gly Glu Leu	260	265	270
Leu Gly Thr Thr Pro Leu Leu Ala Val Asn Ile Cys Lys Phe Lys Gly	275	280	285
Glu Val Ile Ala Lys Asn Gly Asp Val Arg Ser Tyr Arg Met Asp Met	290	295	300
Glu Ile Thr Asn Thr Asp Gly Thr Pro Ile Asp Pro Thr Glu Asp Thr	305	310	315 320
Pro Gly Pro Ile Gly Ser Pro Asp Phe Gln Gly Ile Leu Phe Gly Val	325	330	335
Ala Ser Gln Arg Asn Lys Asn Glu Gln Asn Pro Ala Thr Arg Ala His	340	345	350
Glu Ala Ile Ile Asn Thr Gly Gly Asp His Leu Cys Pro Gln Ile Ser	355	360	365

Ser Ser Glu Ile Tyr Leu Thr Ser Pro Asn Ile Leu Arg Cys Thr Asn
370 375 380

Pro Gln Pro Leu Pro Gln Ser Gly Leu Arg Gly Thr Ile Leu Ile Arg
385 390 395 400

Ser Asp Asn Gly His Cys His Asp Met Val Gly Thr Ser Pro Thr Thr
405 410 415

Pro Thr Trp Pro Gln Gln Trp Arg Arg Cys Ser Arg Gly Ser Asn Cys
420 425 430

Cys Ser Ser Gly His Arg Tyr Pro Val Pro Val Val Met Asn Arg Val
435 440 445

Thr Trp Ile Val Leu Ser His Lys Ser Gly Phe Ser Thr Ser Thr Arg
450 455 460

Lys Leu Pro Gln Leu Asn Leu Arg Trp Pro Leu Ile Arg Phe Ile Asn
465 470 475 480

Pro Asp Thr Gly Arg Val Leu Phe Glu Ala Arg Leu His Lys Gln Gly
485 490 495

Phe Ile Thr Val Ala His Thr Gly Asp Asn Pro Ile Val Met Pro Pro
500 505 510

Asn Gly Tyr Phe Arg Phe Glu Ala Trp Val Asn Gln Phe Tyr Ser Leu
515 520 525

Ala Pro Val Gly Thr Gly Lys Gly Arg Arg Arg Val Gln
530 535 540

<210> 12

<211> 1638

<212> DNA

<213> Hu/NLV/Kashiwa 645/1999/JP

<400> 12

atgatgatgg cgtctaagga cgccccaaca aacatggatg gcaccagtgg igccggccag 60
ctggtaccag aggcaaatac agctgagcct atatcaatgg agcctgtggc iggggcagca 120
acagctgccg caaccgtgg ccaagttaat atgattgacc cctggataat gaataattat 180
gtgcaagccc ctcaaggta atttaccata tcgcctaata acacaccagg tgatatitig 240
tttgatctac aattagcccc tcatctcaal cctttcttat ccatttggc ccaatgtat 300
aacggttggg ttggcaatat gaaagtgaag gtcctattgg ctggtaatgc cttcacggct 360

gglaaaataa tcattagttg cataccccct ggctttgctg cgcaaaacat ttctatcgct 420
 caggccacaa tgttccccca cgttatagct gatgttaggg ttttgaacc tattgaggtg 480
 ccatlgggaag atgtgaggaa tgtgctgttc cataacaatg acaacgcacc aacctgagg 540
 ttgggtgtgca tgcctacac ccccttgcga gccagtggtg gctcatctgg aactgacct 600
 ttgtgtattg ctgggcgtgt tctgacatgc ccaagccctg actttagctt ctatttcttg 660
 gttccccca atgtagagca aaagactaaa ccttttagtg tcccaaatct tccactgaat 720
 accctttcaa attcaagagt ccttctctta attaaatcaa tgatggtaic cagagacct 780
 gggcagatgg ttacagttca aaacggtagg gtcacctg atgggcaact gcaaggcacc 840
 acgcccacat cagctagcca gctgtgcaaa atcagaggca gtgtcttcca tgctaattgt 900
 ggggaatggat ataacctaac tgaattggat gggagcccat accatgcttt tgagagccct 960
 gcgccaatag ggtttcctga tctaggigaa tgtgattggc acatggaggc ctcccctacc 1020
 acccaattca atactgggtga tgttataaaa caaattaatg tcaacaaga atcagcattt 1080
 gctccccacc ttgglacct acaagcagat ggcttgagt atgtgagtgt caacactaac 1140
 atgatagcca aattgggatg ggtgtcacc gtcagtgtg gacatagagg agatgtcgat 1200
 ccgtgggtca ttccacgata tgggtcgact ttgaccgagg ccgccaatt agcccccca 1260
 atatatcccc caggttttgg tgaggccatt gtgttttca tgtcagattt tcctatagcc 1320
 catggtacca atggcttgag tgtgccttgc accatacccc aagaatttgt caccatttt 1380
 gtcaatgaac aggcccctac tagaggggaa gcagccctac tgcatattt agaccctgat 1440
 acccatagaa atcttgggtga gtttaaatla taccctgagg ggttcatgac gtgtgtgct 1500
 aattccagt gacatggctc acaaaccttc ccaatcaatg gtgttttgt ttttgttcc 1560
 tgggtttcca gattctatca gttaaagcct gtgggaacag ccggcccgcc ttgtaggctt 1620
 ggcatcagaa gatcataa 1638

<210> 13

<211> 1593

<212> DNA

<213> Hu/NLV/Seto 124/1989/JP

<400> 13

atgatgatgg cgtctaagga cgctacgtca agcgtggatg gcgccagtgg cgctggtcag 60
 ttggtaccgg aggttaatgc ttctgacct ctgcaatgg atctgtggc gggttcttca 120
 acagcagttg caactgctgg gcaagttaac cctattgacc ctggataat caataacttt 180
 gtcagagctc ccaagggtga atttactatt tctccaaata atacccecg tgggtgtttg 240
 ttgatttga gctaggccc tcatcttaat ccttcttgt tacattgtc acaaatgtat 300
 aatggctggg ttggcaacat gagagttagg attatgctgg ctgglaatgc atttactgca 360
 ggcaaaatta tagtttcttg catacctcct ggctttggct ccataatct tactatagca 420
 caagcaactc tcttcccgca tgtgattgct gatgttagga cttagaccc aattgaagta 480
 ccttgggaag atgtaaggaa tgttctcttt cataataatg atagaaatca acaaacctatg 540
 cgccttgtgt gtatgcttta taccctctc cgcactggtg gcggtacagg tgattctttt 600
 glggttgtag ggcgagtcac gactgtcct agccccgatt tcaatttctt gttcttgggt 660
 cctccacag tggacagaa gactaggcct ttcacctcc caaatttacc gctgagttct 720
 ttgtcaaatt cagtgctcc tcttccaatt agtggcatgg gtatttctcc agacaatgtt 780
 cagagtgtgc agtttcaaaa tggccgatgt accttagacg ggcttctgt tggtagcacc 840
 ccagtttccc tctccacgt tgctaagata aggggcactt ctaatggtac tgtgatcaat 900
 ctacccgaat tggatggcac ccccttccac ccttttgaag gccctgcccc tattggtttt 960

ccagatcttg gggctgtga ttggcatatt aatatgacac aatttgggca tccagtcag 1020
 actcaataig atgtagatag caccocccgac accttcgtcc ctacttagg tccaatccag 1080
 gcgaatggca ttggtagigg caaciatatt ggtgttctta gctgggtctc ccccccata 1140
 catccatctg gctctcaagt tgaatctcgg aagatcccca actatgggtc tagcatcaca 1200
 gaggcaaccc atctagctcc ctctgtctat cctcctggct ttggagaggt gttagtcttt 1260
 ttcatgtcaa agataacctg tccgtgtgct tatagtctgc cctgtttact gccacaagaa 1320
 tataatcac acctcgcaag tgaacaagcc cccactgttg gtgaggccgc ctgtctccac 1380
 tatgttgacc ctgacacggg cgggactctt ggggagtta aggcttacc tgatggtttc 1440
 ctaacctgtg tccctaacgg ggccagctcg ggcccacaac aactaccaat caatggagtc 1500
 tttgtctttg tttcatgggt gtccagattt tatcagttaa agcctgtggg aactgccagt 1560
 tcggcaagag gtaggcttgg tttgcgccga taa 1593

<210> 14

<211> 1641

<212> DNA

<213> Hu/NLV/Funabashi 258/1996/JP

<400> 14

atgatgatgg cgtctaagga cggccctcaa agcgctgatg gcgcaagcgg cgcaggctcaa 60
 ctggtgccgg aggttaatac agctgacccc ttacccatgg aaccctggc tggccaaca 120
 acagccgtag cactgctgg gcaagttaat atgattgac cctggattgt taataatttt 180
 gtccagtcac cacaaggiga gtttacaatt tccctaata ataccccggt tgataatttg 240
 ttgtatttac aattaggctc acatctaaac ctttcttgt cacatctgtc ccaaagtgtat 300
 aatggctggg ttggaacat gagagttagg attctccttg ctgggaatgc attctcagct 360
 ggaaagatta tagtttgttg tgcctccctt ggctttacat ctctctctc caccatagct 420
 caggctacat tgtttcccca tgtgattgct gatgtgagaa ccttgaacc aatagaaatg 480
 cccctcgagg atgtacgcaa tgcctctat cacaccaatg ataataacc aacaatgcgg 540
 ttggtgtgta tgcgtacac gccgtccgc actggtgggg ggtctggtaa ttctgattct 600
 ttgtgtgttg ctggcagggt gctcacggcc cctagtagcg acttcagttt cttgttctt 660
 gtcccgcta ccatagaaca gaagactcgg gcttttactg tgcctaata ccccttgcga 720
 accttgtcca attctaggtt tcttccctc atccaggga tgaattctgt tctgacgca 780
 tctcaagtgg tccaattcca aaatggacgt tgcctcatag atggtaact cctaggcact 840
 acaccgcta catcaggaca gctgttcaga gtaagaggaa agataaatca gggagcccg 900
 acgtcaacc tcacagaggt ggaatggcaa ccattcatgg catttgatc cctgcacct 960
 gtgggttcc cggatttttg aaaatgtgat tggcacatga gaatcagcaa aaccccaat 1020
 aacacaagct caggtagacc catgcgcagt gtcagcgtgc aaaccaatgt gcagggtttt 1080
 gtgccacacc taggaagtat acagtgtgat gaagtgttca accacccac aggtgactac 1140
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1641

<210> 15

<211> 1635

<212> DNA

<213> Hu/NLV/Chiba 407/1987/JP

<400> 15

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ggtgtccgta gataa 1635

<210> 16

<211> 1620

<212> DNA

<213> Hu/NLV/Narita 104/1997/JP

<400> 16

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 gtagcgggcc aacaaaatgt aattgacccc tggattagaa ataattttgt acaagcccct 180
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 ttgggcccctg atttgaaccc ctacctttct catttggcca gaagtataca tggttatgca 300
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 actggccgc atgatttggg tatccccccc aatggttact ttgatttga ctctgggtc 1560
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<210> 17

<211> 1647

<212> DNA

<213> Hu/NLV/Sanbu 809/1998/JP

<400> 17

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 ctactgtgtc agcaaaacat aattgatccc tggattatga ataattttgt gcaagcacct 180
 ggtggtttagt ttacagtgtc ccttaggaat tcccctgggt aagtgttctt taatttggaa 240
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aggtttccag tgccgattga gtccttgcac accagcccaa ctgagaatat tgtgtccag 780
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 ggatatttta ggtttgaatc ttgggtgaac ccttttata cacttgcccc catgggaact 1620
 gggaacgggc gtagaaggat tcaataa 1647

<210> 18

<211> 1623

<212> DNA

<213> Hu/NLV/Ichikawa 754/1998/JP

<400> 18

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 gtcactggcc aaactaatat aatagacccc tggattagaa ctaattttgt ccaagcccc 180
 aatggatgaat ttacagtctt ccctagaaat tcccctggag agatattggt caatttggag 240
 ttgggtccag aactgaaccc ttatctggca cathtagcta ggaatgataa tggttatgag 300
 ggtggtatgg aggtgcaagt gatgctcgcg gggaacgcgt tcaactgtgg caagatcatc 360
 ttgccgccg tgccacctta cttccagtg gaaaatctta gcccttccca aataacaatg 420
 ttcccacatg tgatcatcga tgcagaacc ttggaacctg tattactccc aatgcctgat 480
 gtcagaagca cccctttcca cttaatacaa aaagatgagc ctaagatgag acttgttgcc 540
 atgctttaca cccccctcg ttctaatggt tctggtgacg acgtttcac cgtctcatgt 600
 aggatcctca ctaggccctc cctgaattt gattttacat atttggtagc accaacagta 660
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 agattccctc tctctattga tgaatggtc accagcccaa atgagtcct agttgttcag 780
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 agtactagca cagggcctgt tgtggttcca gccaatggct atttcaaatt tgattcctgg 1560
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<210> 19

<211> 1608

<212> DNA

<213> Hu/NLV/Chitta 1876/1996/JP

<400> 19

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 ctcaccggcc aaaacaatat tatagacccc tggattagat taaattttgt gcaggctccc 180
 aatggagagt tcacggtttc accccgcaac tcaccggggg aagtcctatt aaatttggaa 240
 ttaggccccg aactaaatcc atacctagca caccitttcta gaatgtataa tggttatgca 300
 ggtggggttg aggtgcaagt actactggct gggaatgcgt tcacagctgg aaaattggig 360
 tttgccgcag ttccccccta tttccatta gaaaacataa gccctggta gataactatg 420
 tttctcatg taattattga tgttaggact ttagaaccag ttttgttgc ccttctgat 480
 gtttaggaata atttctttca ttataatcag cagaatgaac cgaggatgag actcgtagca 540
 atgctttata ctctcttag atctaattgt tctgggtgat atgtatttac tgtctctgc 600
 aggggtgcta ccgaccttc ccctgatttt gattttaatt acttggctcc ccctaccctt 660
 gaatctaaaa cttaaccctt cacactccct atcttgacta taggggagtt aaccaactcc 720
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 ccgattgttg taccagctaa tggttacttc aggtttgata ctgggttcaa tcaattctat 1560
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<210> 20

<211> 1629

<212> DNA

<213> Hu/NLV/Kashiwa 47/1997/JP

<400> 20

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gtggcgggac aaaccaacat aattgacccc tggataagaa caaattttgt acaagccccc 180
aatggagagt ttacagtgtc accaagaaat tcccctggag aaattttatt aaatttagaa 240
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cttccccatt tgattgtaga tgttaggact ttggaacctt ttatgacacc ctgcctgat 480
gttaggaatg tgttctatca ttttaataat caacctcaac ctagaatgag gttagtggct 540
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cggttcccca ttccaatcga gcaattgtat acggtcccaa atgaaaccaa tgttgtccag 780
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tcttgggtta atcaatttta ctacatgcc cccatgggaa ctggcaatgg gcgtagaaga 1620
attcagtaa 1629

<210> 21

<211> 1653

<212> DNA

<213> Hu/NLV/Mie 7k/1994/JP

<400> 21

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gttctggccc agcaaaatat aattgacccc tggattagag aaaattttgt ccaagcaca 180
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tgtcccaatg tgattgttga tgtgagacaa cttgaaccag tgcctctgcc cctccctgat 480
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<210> 22

<211> 1626

<212> DNA

<213> Hu/NLV/Osaka 10-25/1999/JP

<400> 22

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 ggcggcatgg aagtgcacaa tgtgtctggc gggaatgcgt tcacagcagg caaaatcata 360
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